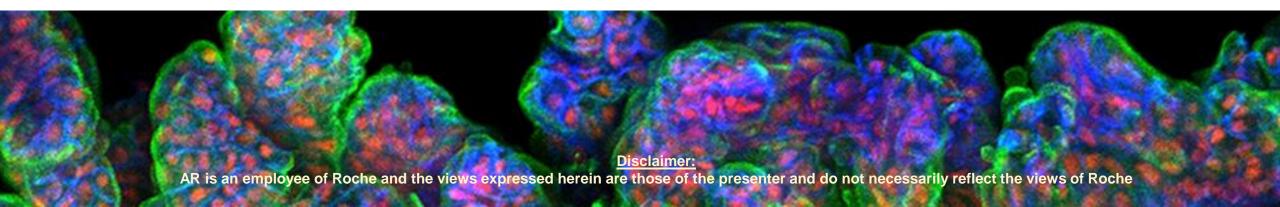


Novel human cell models in drug development: How 3D, Organoids & Organs on Chips can improve and renew current paths - and our vision for the future

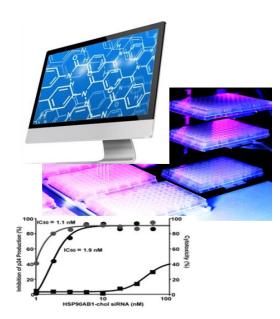
Prof. Adrian Roth, PhD

Principal Scientific Director Personalized Healthcare Safety, Pharma Clinical Development Hoffmann-La Roche Ltd, Basel, Switzerland

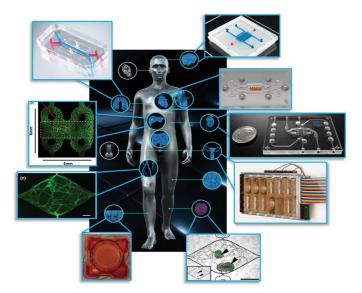


Efforts in Pharmaceutical Industry aiming at increasing Quality & Speed of Drug Development – while reducing Cost & Animal use

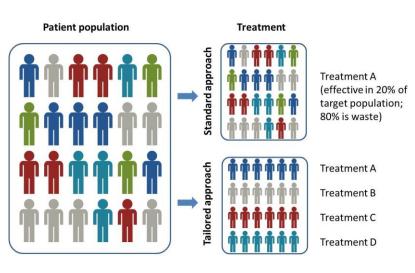
Fail early – fail fast



In silico design, lab automation, Al-powered algorithms Organoids & "Organs on Chips"

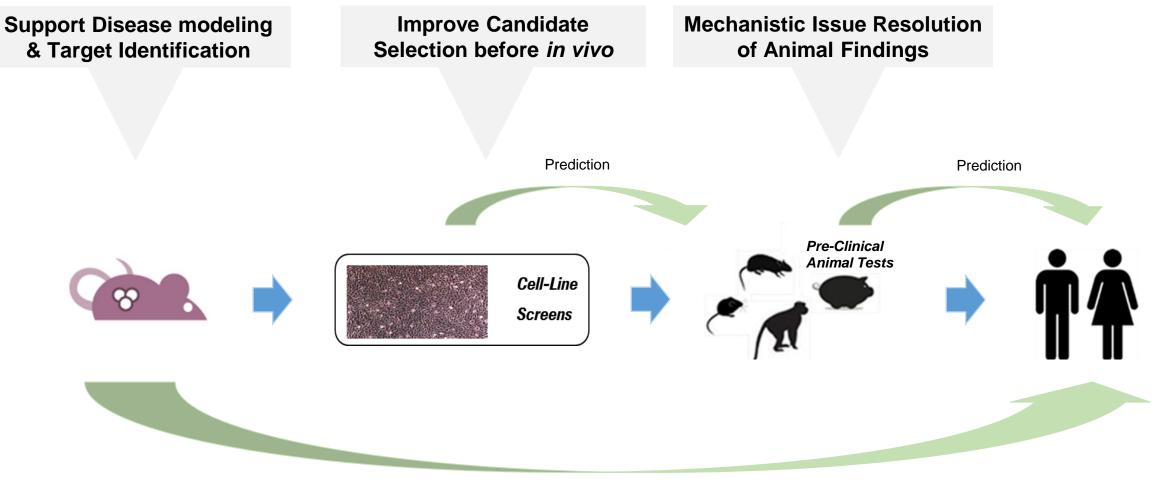


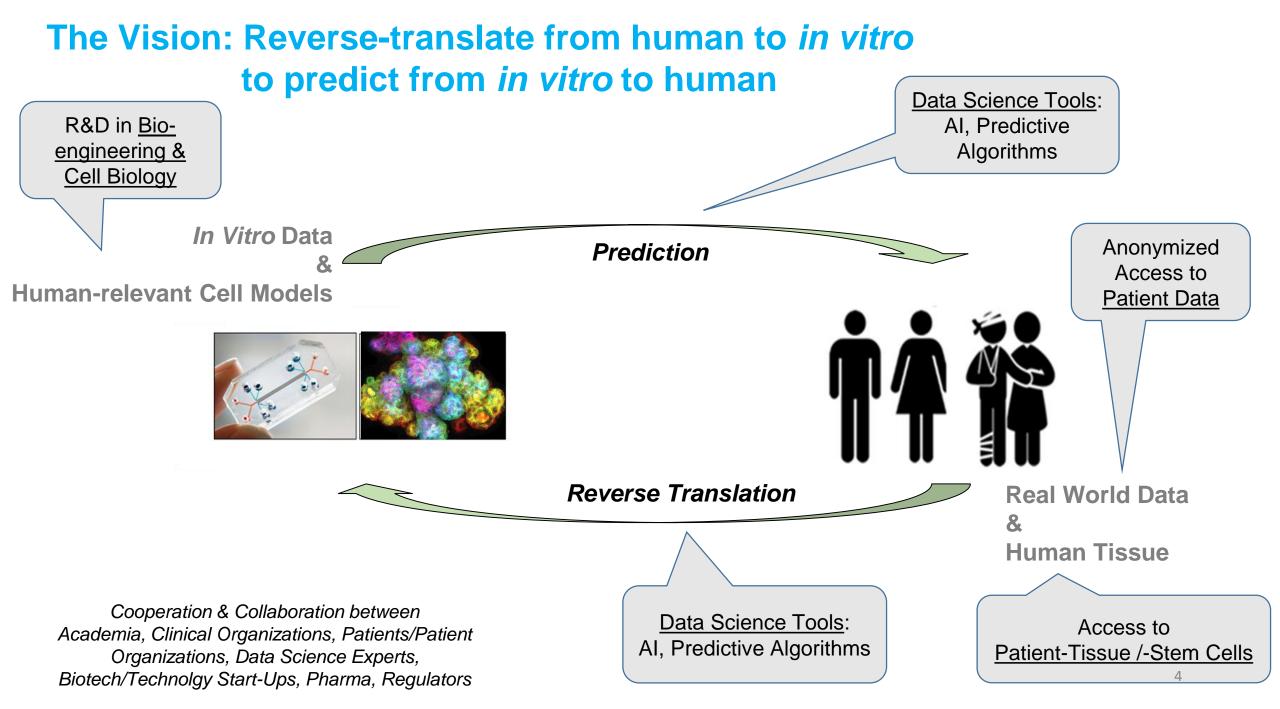
Advanced human cell models that better recapitulate human physiology "Real World Data" & Genetics



Personalized Medicine: Understand & predict human variability – optimize benefit/risk 2

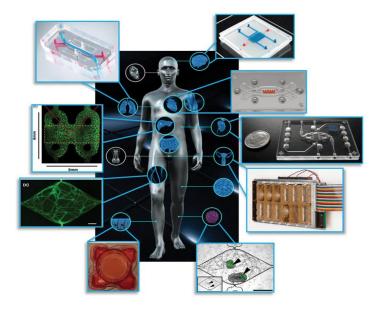
Advanced Human Cell Models today





Advanced human cell models in Pharmaceutical Drug Development: Where are we today ?

- A burst of different approaches some with more, some with less potential for application
- Not fully clear yet which cells, which materials, which setups, which assays or which endpoints are «the right ones»
- A lot of overviews & strategies published yet, best and most convincing way for adoption is to demonstrate compelling evidence for defined <u>use cases</u>
- Broad industry adoption is slow reasons may include
 - high upfront investments
 - complex technical setting
 - lack of robustness
 - lack of clear superiority over existing models
 - unclear translational potential to the human situation



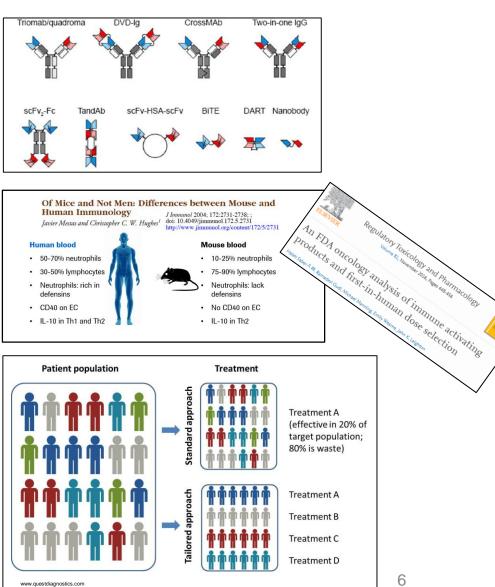
Opportunities and challenges with microphysiological systems: a pharma end-user perspective

Lorna Ewart¹ and Adrian Roth²

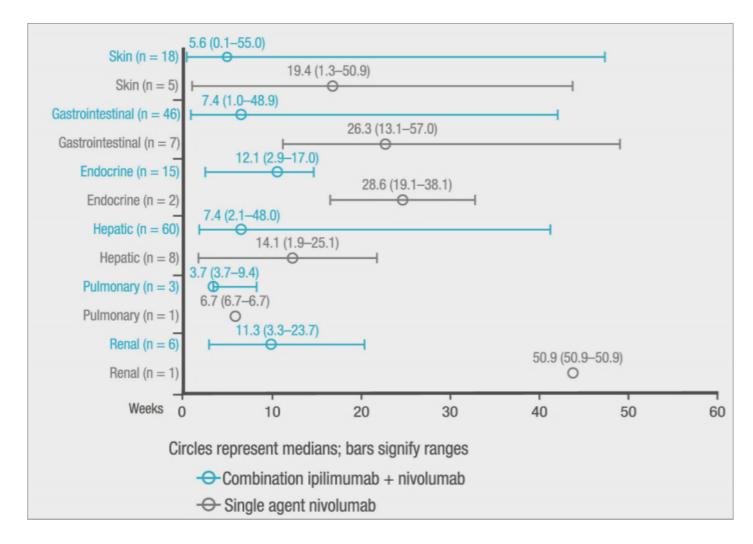
Using human-relevant, translational in vitro models is widely considered to reduce attrition during drug discovery and development. Despite this, the adoption of models based on microphysiological systems — organs-on-chips or organoids — by pharma companies is moderate at best, and realizing the full potential of these models will need greater collaboration between stakeholders.

What drives application of Advanced Human Tissue Models in Pharmaceutical Drug Development?

- Shift in portfolio from small to complex, engineered molecules that often have multiple targets
- Molecules often do not cross react with any pre-clinical species (not even primates)
- **Target(s) & pathway(s)** are not adequately represented in any animal species (i.e. **immune-related**)
- Additional upcoming Challenges:
 - Need for assessing safety & efficacy in children
 - Need for assessing safety & efficacy in different ethnicities
 - Sometimes small patient population
 - Goal to increase benefit/risk, ie strive for more personalized medicine
- Conventional pre-clinical *in vivo* testing may not be relevant or simply <u>not possible</u>
- Urgent need for novel tools to assess the pharmacology & toxicology of these new types of drug candidates



Cancer Immunotherapy: Clinical Toxicities of Immune-engaging Antibodies

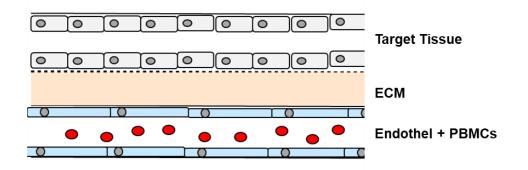


Time to onset of grade 3–4 treatment-related selected adverse events (AEs)

Ipilimumab:Anti-CTLA-4 Nivolumab: Anti-PD-1

Annals of Oncology Volume 28, Supplement 4, July 2017, Pages iv119iv142 Suite of Human Models established to address "on-target, off-tumor" safety liabilities of bi-specific antibodies

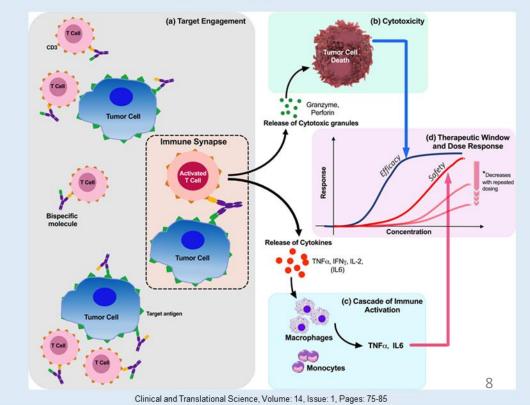
- Primary human tissue, polarized where required
- **PBMCs** in flow
- All tissue/cell components donor/HLAmatched



Current conventional 2D cell systems lack essential cellular, biochemical, and biophysical factors found in the native organ

Goal to create models that are able to recreate

- Drug induced activation of immune cells
- Initiation of downstream events including cell migration to site of action
- Measurable effects at site of action that allow drawing conclusions on drug molecule properties, desired & undesired effects, relevance to patient population



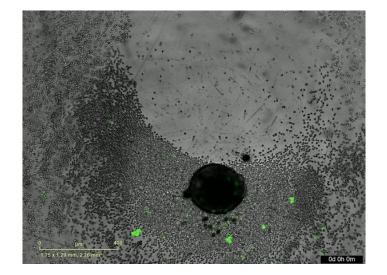
Predict immune-related toxicities early on by detecting key events using advanced human cell systems

Immune cell activation

Targeting healthy tissue

R Villenave

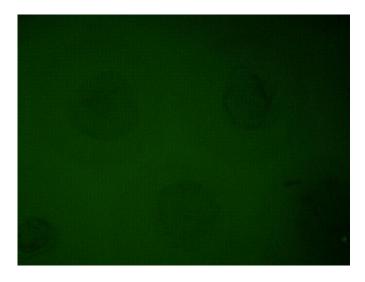
Recruitment of blood leukocytes under flow following TNF-a stimulation



P Godoy/E Breous-Nystrom

Leukocyte infiltration into hepatic spheroids following anti-PD-1 nivolumab treatment

Destruction of healthy tissue



N Gjorevski/M Bscheider

Chemotherapy-induced apoptosis of intestinal organoids

-> Use relevant cell types, readouts & and known relevant positive and negative controls to validate models for specific context of use

Ask the right questions with the right models

From single to multi-organ model approach – focused questions or broad vital functions for de-risking

Cytokinemediated toxicities

- Rather immediate or short-term effects
- Can be secondary to tumor lysis
- Involve the vasculature and can affect many tissues and organs

On-target / off-tumor toxicities

- Typically restricted to tissues where target is expressed
- Less of a concern if safety window large enough due to significantly higher expression in tumor

Off-target / offtumor toxicities

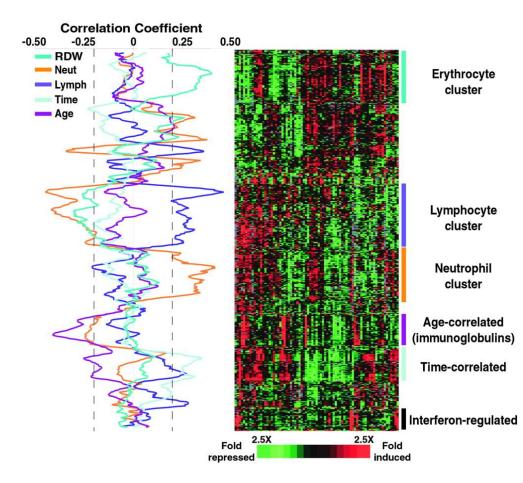
- Difficult to predict as can hit any cell type of the human body
- Can be best addressed with a thorough analysis of off-target peptides

Autoimmunity, infections, inflammation

- Result from an impairment of the immune system
- Can take various forms but could involve similar mode of actions
- Require lymphoid responses

Specifically relevant for T cell engaging molecules and adoptive cell therapies (e.g. Bispecifics and CAR-T)

Example for technical challenges to tackle: Factors affecting blood donor variability



Variation in gene expression patterns in human blood. Whole blood was drawn from 75 healthy volunteers.

- Genetic diversity
- Environmental factors: immunizations, nutrition, latent infections
- Technical factors: Cryopreservation techniques, buffers & media
- Sample composition cellular subsets



Human immunocompetent Organ-on-Chip platforms allow safety profiling of tumor-targeted T-cell bispecific antibodies

S Jordan Kerns, Chaitra Belgur, Debora Petropolis, Marianne Kanellias, Riccardo Barrile, Johannes Sam, Tina Weinzierl, Tanja Fauti, Anne Freimoser-Grundschober, Jan Eckmann, Carina Hage, Martina Geiger, Patrick Ray Ng, William Tien-Street, Dimitris V Manatakis, Virginie Micallef, Regine Gerard, Michael Bscheider, Ekaterina Breous-Nystrom, Anneliese Schneider, Anna Maria Giusti, Cristina Bertinetti-Lapatki, Heather Shannon Grant, Adrian B Roth, Geraldine A Hamilton, Thomas Singer, Katia Karalis, Annie Moisan, Peter Bruenker, Christian Klein, Marina Bacac, Nikolce Gjorevski [©], Lauriane Cabon [©]

Emulate Inc, United States; Emulate Inc., United States; Roche pRED, Switzerland; Roche pRED, Germany

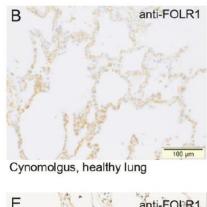
Research Article · Aug 11, 2021

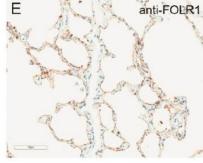
Example: TCB targeting CD3 & FOLR1

Pre-clinical Safety Assessment:

- efficacious in human breast cancer patient-derived xenograft models
- severe on-target toxicity in the lung of cynomolgus monkey
 - *leukocytic infiltrates in lung tissue indicative of immune mediated toxicity*

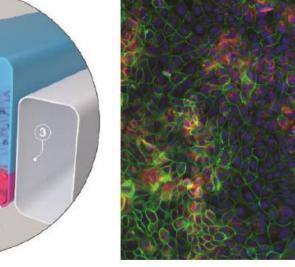
FOLR1 Nucleus E-cadherin





Human, healthy lung

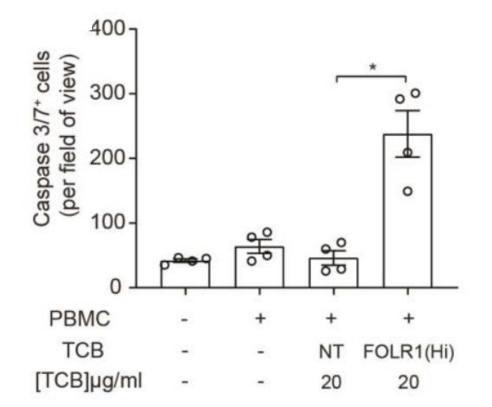
1. Top Channel
2. Human Epithelial Cells
3. Vacuum Channel
4. Porous membrane
5. Human Endothelial Cells
6. Bottom Channel





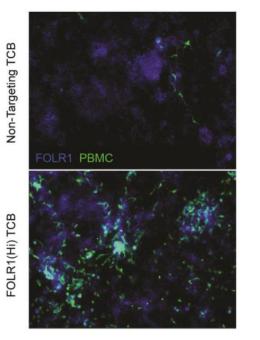
Example: TCB targeting CD3 & FOLR1

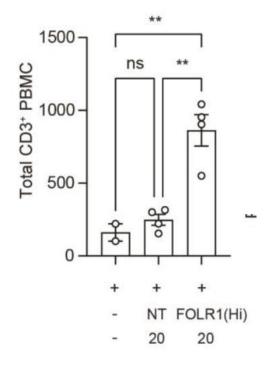
Apoptotic cells collected on live chips



PBMC	Peripheral Blood Mononuclear Cells
TCB NT	Non Targeting TCB
TCB FOLR1(HI)	FOLR1-targeting TCB

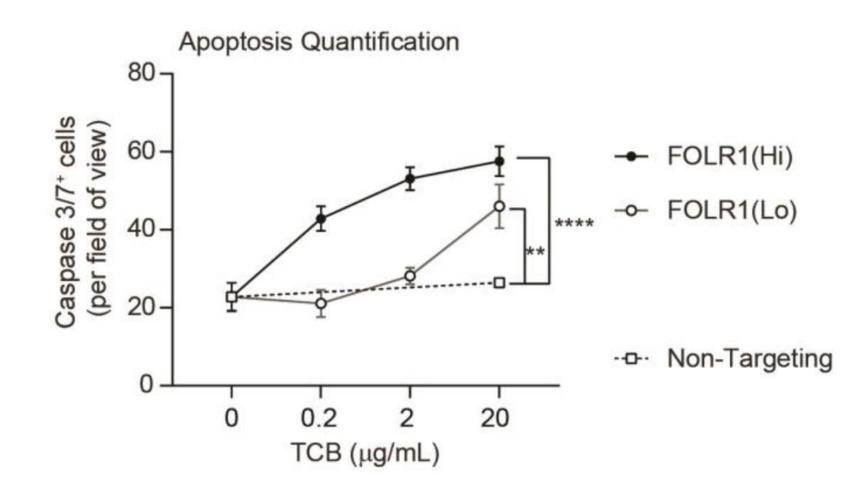
PBMC accumulation at sites of target expression consist primarily of CD3-T cells





Example: TCB targeting CD3 & FOLR1

Model leveraged to identify a safer molecule





Lab on a Chip

PAPER

View Article Online View Journal | View Issue



Cite this: Lab Chip, 2020, 20, 3365

Neutrophilic infiltration in organ-on-a-chip model of tissue inflammation[†]

Nikolce Gjorevski, ⁽¹⁾* Blandine Avignon, Régine Gérard, Lauriane Cabon, ⁽¹⁾ Adrian B. Roth, Michael Bscheider and Annie Moisan ⁽²⁾*

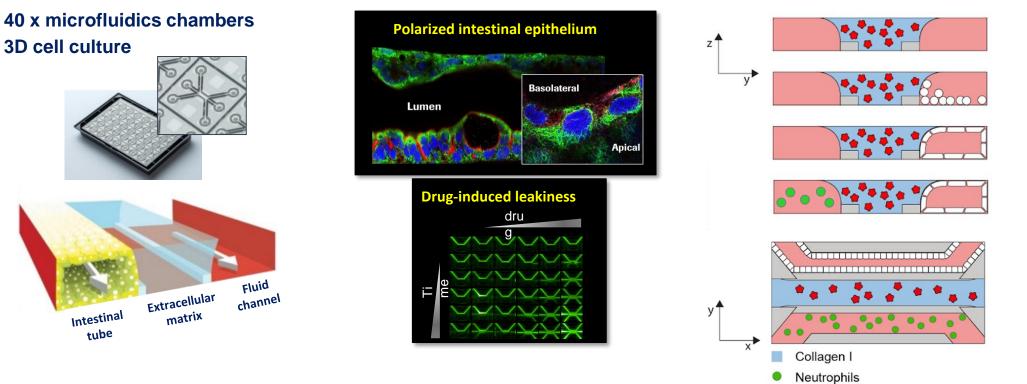
Neutrophilic infiltration in a gut-on-a-chip model of intestinal inflammation



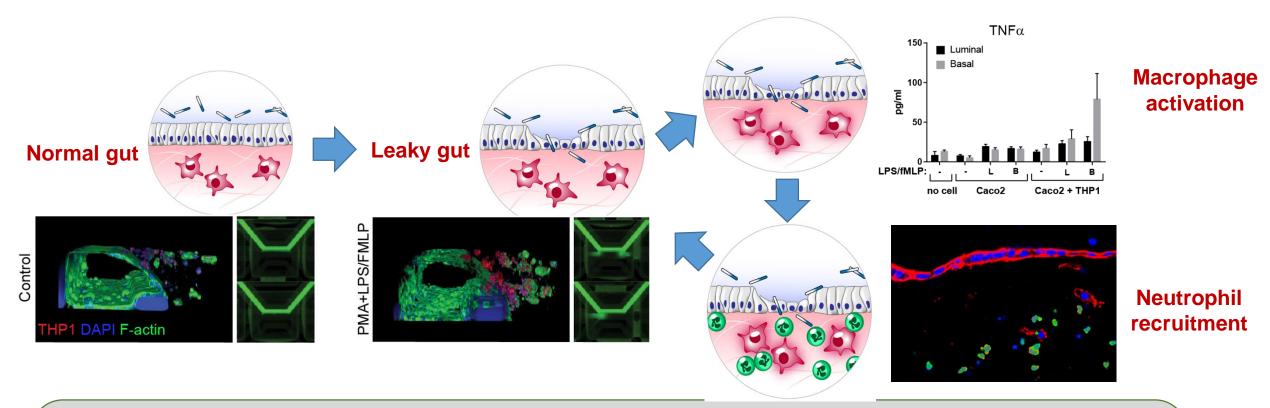
https://www.mimetas.com/en/3d-cell-migration/

THP1

□ Intestinal epithelial cells



Neutrophilic infiltration in a gut-on-achip model of intestinal inflammation



- Tri-culture of epithelial, resident and infiltrating immune cells, capturing their functions and functional interactions
- Infiltrating neutrophils exacerbate the inflammatory process, leading to epithelial damage
- CaCo-based; Missing stromal and vascular cells
- Adaptive immune cell contribution to inflammation not captured

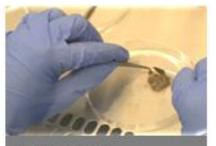
SCIENCE TRANSLATIONAL MEDICINE | RESEARCH ARTICLE

CANCER

Tumor-targeted 4-1BB agonists for combination with T cell bispecific antibodies as off-the-shelf therapy

Christina Claus^{1*}, Claudia Ferrara^{1*}, Wei Xu^{1*}, Johannes Sam¹, Sabine Lang¹, Franziska Uhlenbrock², Rosmarie Albrecht¹, Sylvia Herter¹, Ramona Schlenker¹, Tamara Hüsser¹, Sarah Diggelmann¹, John Challier¹, Ekkehard Mössner¹, Ralf J. Hosse¹, Thomas Hofer¹, Peter Brünker¹, Catherine Joseph³, Jörg Benz³, Philippe Ringler⁴, Henning Stahlberg⁴, Matthias Lauer³, Mario Perro¹, Stanford Chen¹, Christine Küttel¹, Preethi L. Bhavani Mohan¹, Valeria Nicolini¹, Martina Carola Birk¹, Amandine Ongaro¹, Christophe Prince¹, Reto Gianotti¹, Gregory Dugan⁵, Christopher T. Whitlow⁵, Kiran Kumar Solingapuram Sai⁵, David L. Caudell⁵, Armando G. Burgos-Rodriguez⁶, J. Mark Cline⁵, Michael Hettich³, Maurizio Ceppi³, Anna Maria Giusti³, Flavio Crameri³, Wouter Driessen³, Peter N. Morcos⁷, Anne Freimoser-Grundschober¹, Victor Levitsky¹, Maria Amann¹, Sandra Grau-Richards¹, Thomas von Hirschheydt⁸, Stella Tournaviti⁸, Michael Mølhøj⁸, Tanja Fauti¹, Viola Heinzelmann-Schwarz⁹, Volker Teichgräber¹, Sara Colombetti¹, Marina Bacac¹, Alfred Zippelius², Christian Klein¹, Pablo Umaña^{1†} Copyright © 2019 The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works

3D tumor explant culture system



dissection





Baseline

D1

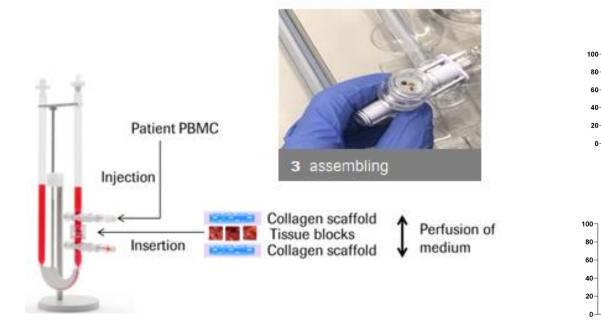
D2

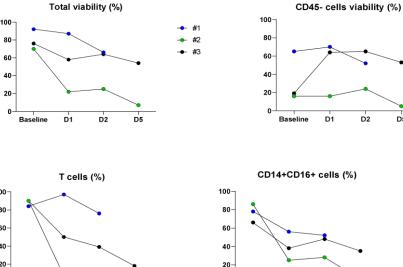
D5

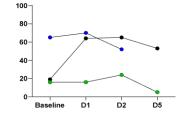
Viability is stable for two days (48h), drops afterwards "Viability drop" equally observed in all cell populations

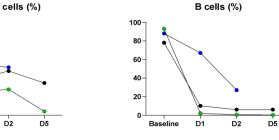
Baseline

D1







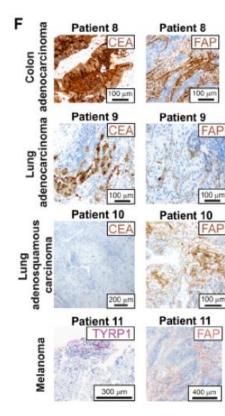


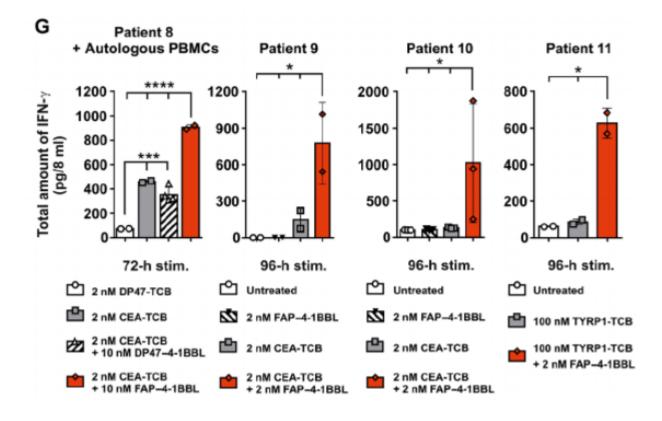
TCB combination with tumor-targeted 4-1BBL agonist

Is natural expression of FAP in human tumor tissues sufficient to provide functional cross-linking for TCBs ?

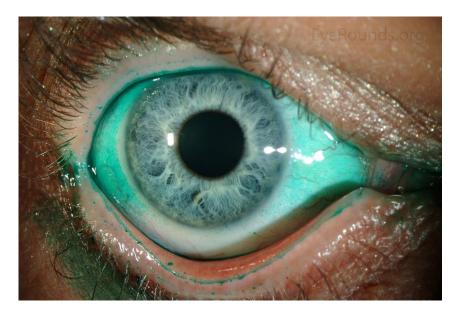
Target (FAP, CEA, or TYRP1) expression in tumor tissue samples

Drug treatment of cultured tumors in 3D system



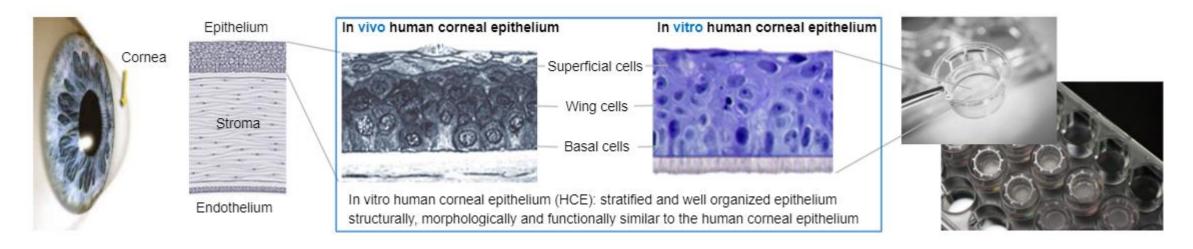


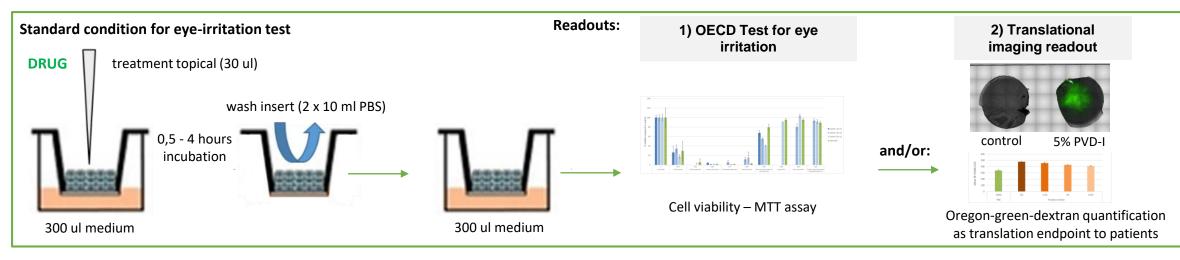
From an OECD qualified test for eye irritation to safety and efficacy modelling for pharmaceutical drugs



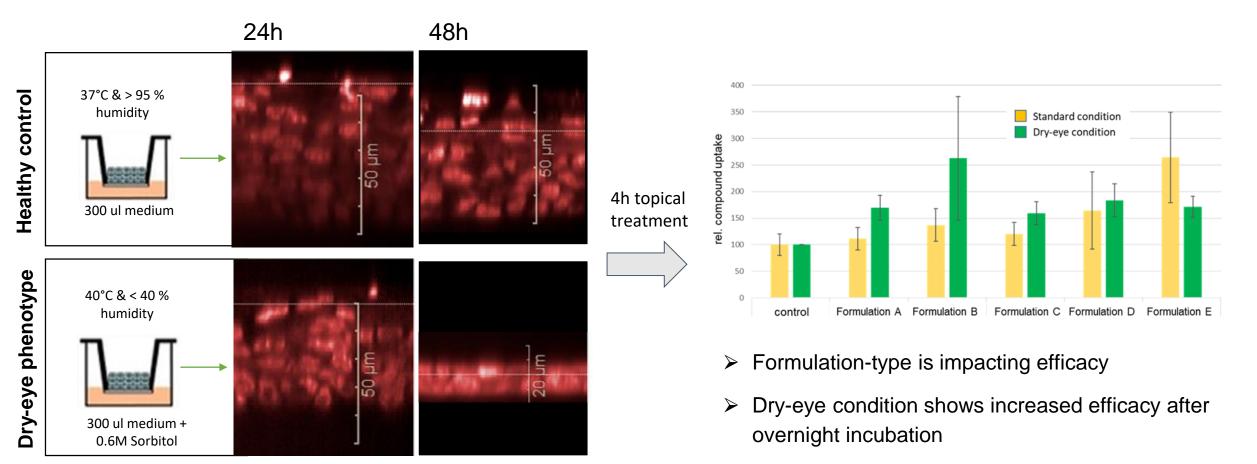
(Karen Dernick, Claudia Korn, Michael Keller, Christian Bucherer, Christoph Ullmer and Stefan Kustermann)

Human 3D Corneal Epithelium as in vitro Model for "Front of the Eye"- Applications and Eye Irritation Tests





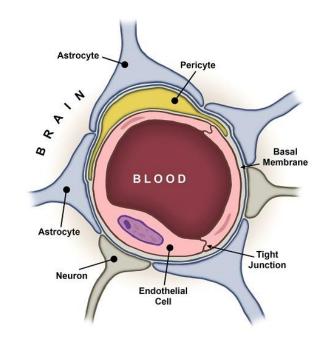
Improved prediction of efficacy using «dry eye disease model»: disease phenotype and formulation drives efficacy



Dry-eye phenotype can be induced in the model and leads to a decrease in the epithelial thickness after 48h • Proposed mechanism: higher uptake of compound due to impaired barrier

A Blood Brain Barrier Model based on Organoids

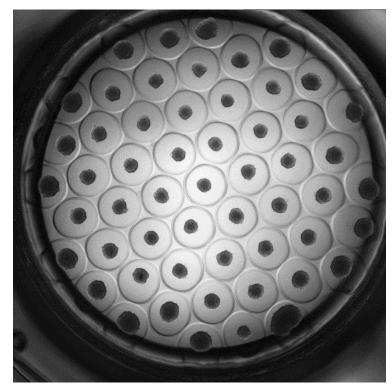
(Roberto Villasenor Solorio)



BBB organoid arrays for high-throughput screening

Imaging frame at spheroid

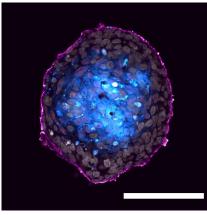
core

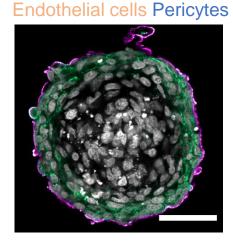


- Up to 3000 organoids per plate
- Highly reproducible size (150 250 mm Ø)
- Compatible with automated microscopy

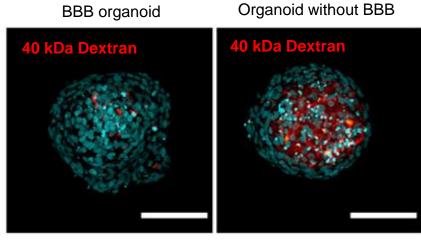
Self-assembly of the human neurovascular unit

Endothelial cells Astrocytes

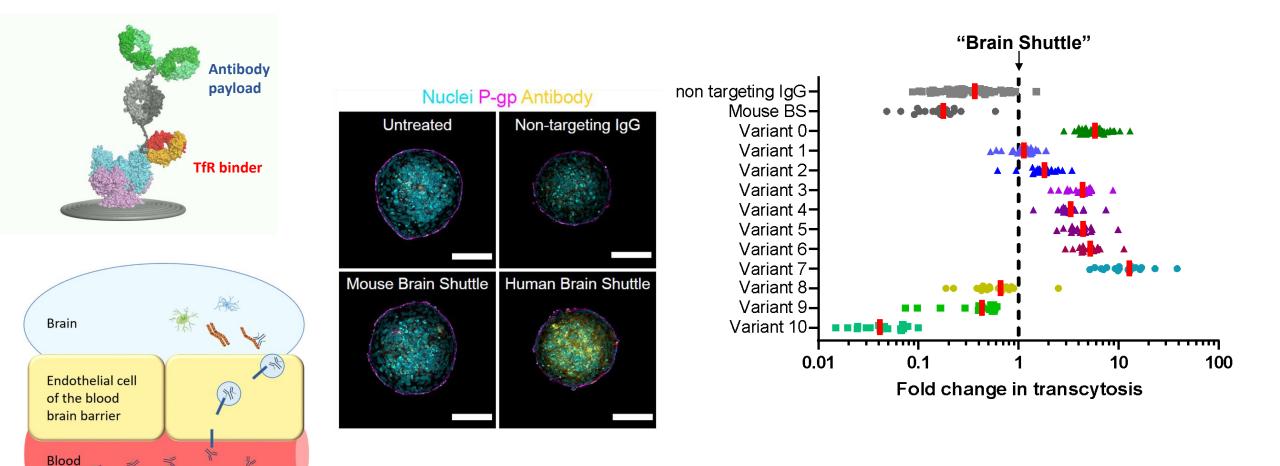




BBB organoids recapitulate barrier properties for biologics



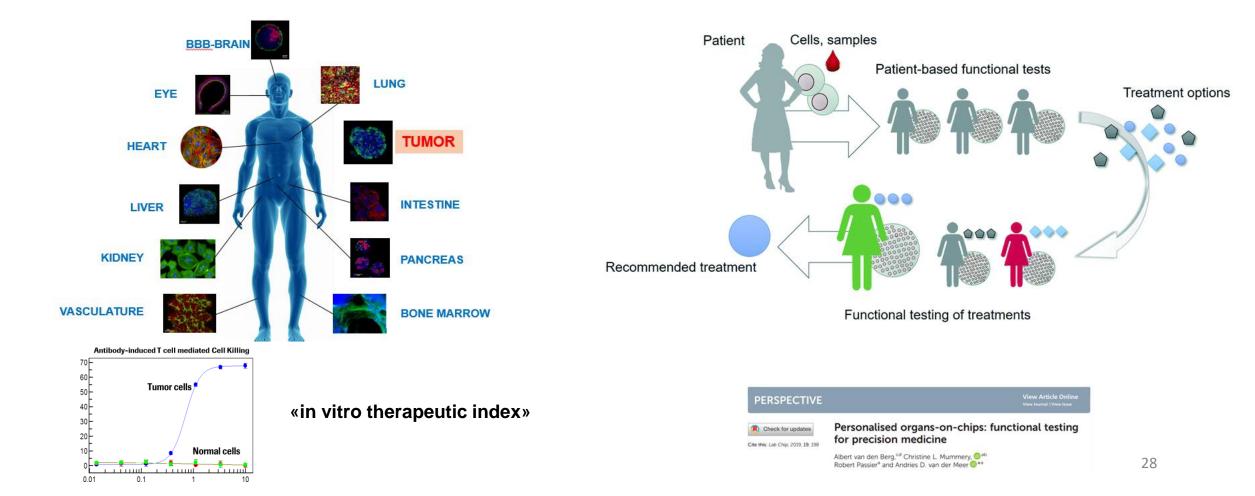
Screening of new modalities for CNS delivery BBB organoids accelerate discovery and optimization cycles



Where do we want to go?

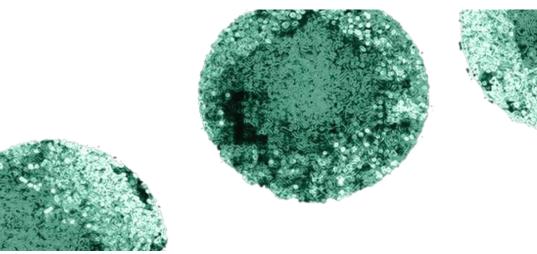
In vitro efficacy & safety assessment for EIH-enabling

Patient-derived models enabling personalizing clinical trials



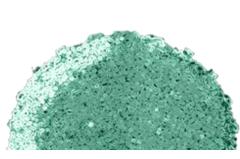
A strong commitment to build on human cell based approaches to support drug development:

Roche Institute for Translational Bioengineering (ITB)

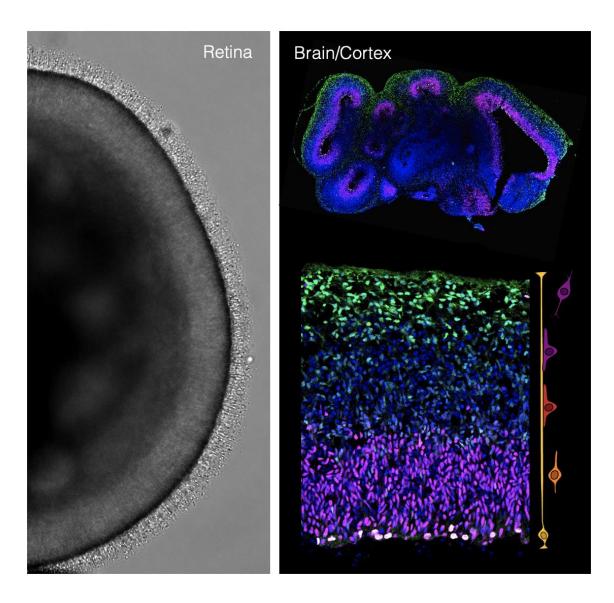


An incubator and catalyst for big, bold ideas and creative organoid technologies that can be translated into drug development for the benefit of patients

Prof M. Lütolf



TRANSLATIONAL BIOENGINEERING 'Organoid Factory': Brain and retina organoid manufacturing



- High-throughput, automated mass production of retinal and brain organoids
- Integrated quality control module (imaging- and AI-based) with organoid sorter
- Collaboration with selected external partners on specific aspects (e.g. organoid sorting technology)
- Explore miniaturization and bioreactor-free culture
- Proof-of-principle results for manufacturing of other organoid systems



Advanced Human Tissue Models in Drug Development: Outlook

- Significant Investments in Academia, Biotech & Pharma have led to a series of encouraging Use Cases that demonstrate the Potential of more complex, physiologically relevant human Cell Models
- Broad Industry Adoption is low due to
 - Immaturity of some of the Systems
 - Complex technical Set-up not suited for Scaling and daily Use
 - > High Investment needed Pletora of different Approaches
 - Biological Relevance not convincingly shown and/or Lack of Superiority over already exisitng Models
- Areas with high unmet Need could help driving Application forward, ie where conventional Models are not an Option (e.g. Immunology), where lean Drug Development Paths are possible (e.g. Rare Diseases), where fast Reacting to an urgent Need is warranted (e.g. COVID)
- Advanced Human Tissue Models not only can significantly improved pre-clinical Development Phases – they could as well become Game Changers for Clinical Development (i.e. Bedside-Bench-Bedside, Personalized Medicine)
- Next to access to Patient Tissue/Cells Access to Patient Data is key for renewing Drug Development Paradigm

Science 17 Sep 2021, Vol 373, Issue 6561, pp. 1304-1306

PERSPECTIVES

MEDICINE

Human microphysiological systems for drug development

Organs-on-chips could be used to assess drug efficacy and support personalized medicine

By Adrian Roth¹ and MPS-WS Berlin 2019²

icrophysiological systems (MPS), such as microfluidic organs-onchips, have rapidly evolved as promising in vitro tools to recapitulate human physiology by recreating key biological processes and disease states. However, their value for drug development is only now becoming interconnections with other organs (4, 5), c They add to the toolbox of assays to identify potential therapeutics for diseases, including COVID-19 (6). These features enable human multi-cell-type systems that can better replicate complex tissue and organ functions than conventional cell culture. Consequently, MPS have gained broader attention as a tool to improve the prediction of human efficacy and potential undesired effects of drugs before patients are exposed to them (7).

clear. MPS combine microsystems engineer-MPS technologies may provide a way to ing with cell biology, yielding cell-culture better understand and address the main models that can display three-dimensional failures of clinical programs: lack of effiarchitecture, multicellular interactions, cacy or unacceptable side effects that are tissue-tissue interfaces, fluid flow, and not predicted in animals or simpler cell organ-level mechanical cues. For example, systems during early preclinical stages. The they can incorporate breathing mechanics key advantage that MPS offer is the creation of human lungs (1), circulating immune of more physiologically relevant human orcells trafficking through perfused microgan-like models that can potentially vield data on drug action that will better transvasculature (2), living microbiotas (3), and

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https://www.science.org/doi/10.1126/science.abc3734



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Shamin Li Tamara Hüsser Inês Silva Steffen Dettling Karen Dernick Claudia Korn Christoph Ullmer Christoph Ullmer Christine Zihlmann Anja Osterwald Christian Bucherer Marlene Juedes Matthias Lütolf Heloise Ragelle



- Emulate
- TissUse
- Mimetas
- SUN Bioscience
- Don Ingber, Jennifer Lewis @Wyss
- Hierlemann Lab @ETHZ
- Hartung Lab @JHopkins
- InSphero
- Loskill Lab @ Fraunhofer
- AlveoliX