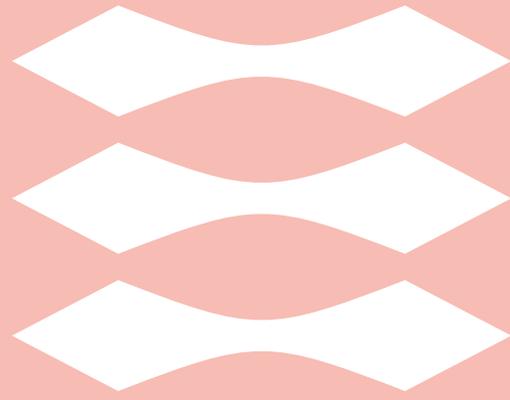


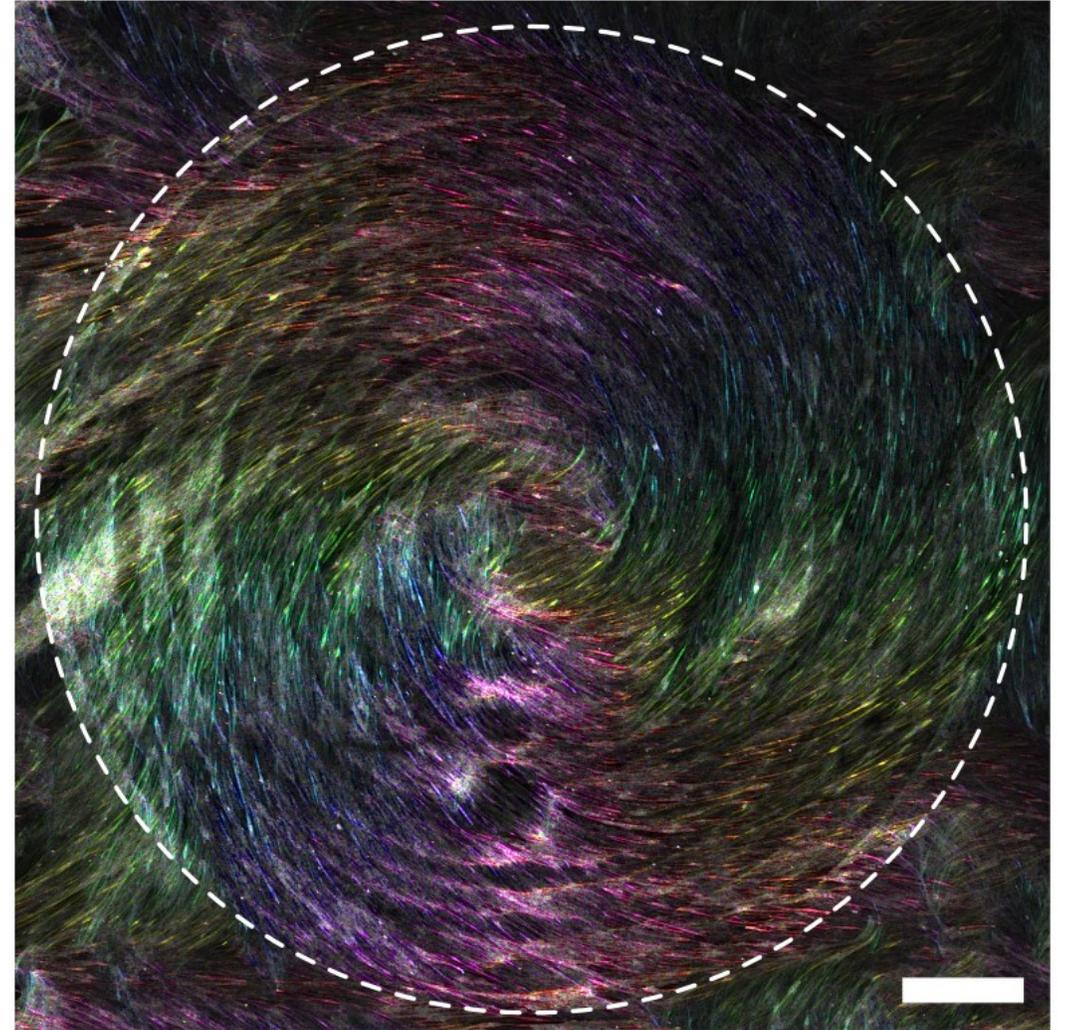
DTU



# Instrumented and 3D printed human tissue models

# Outline

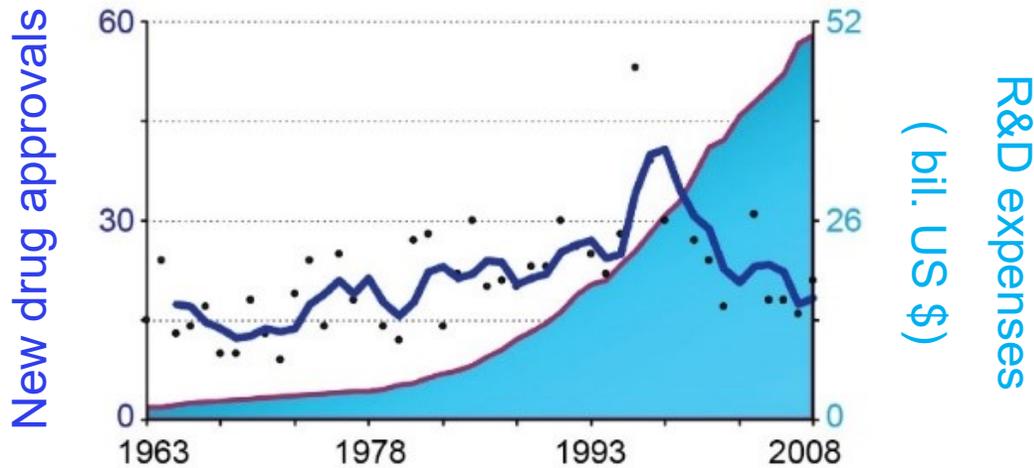
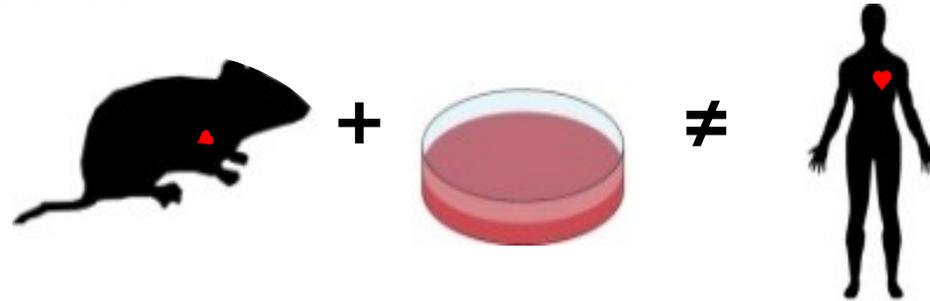
- Engineering human tissue models
- Instrumented cardiac tissue models
  - Multi-material 3D printing
  - Application examples
- Cell-instructive biomaterials
- Embedded Bio-printing
- Summary & Outlook



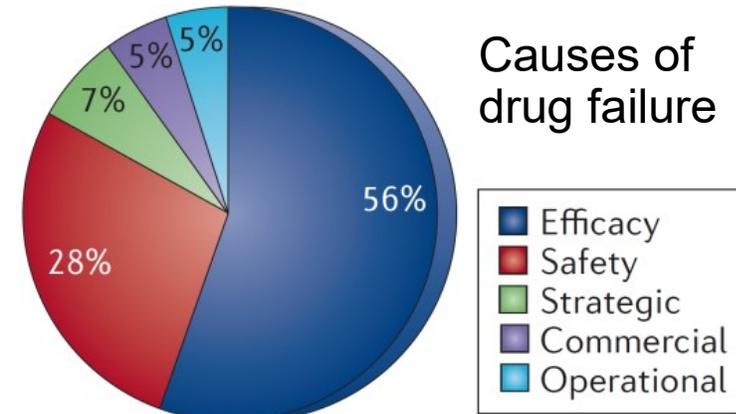
Adapted from SD Cakal et al *biomedical materials* 2022

# Engineering human tissue models

# Old problems



Kaitlin Kl et al *Clin. Pharm. & Therapeutics* 2010



Arrowsmith J, Miller P.

*Nature Reviews Drug Discovery*. 2013

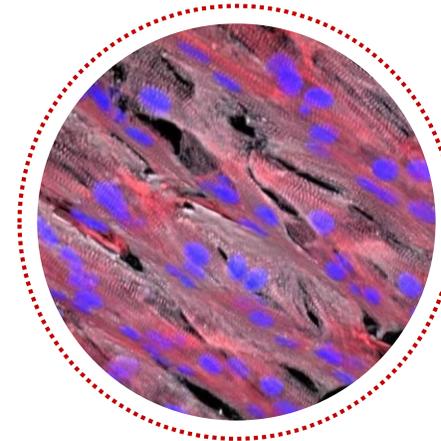
The traditional drug development pipeline is deeply flawed

# New solutions?

## Human iPSC-based models

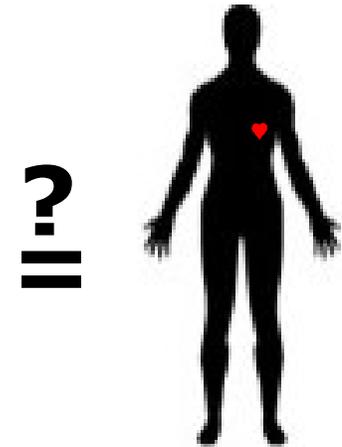
*Organoids*, *organs-on-chips* & *3D bio-printing*, share

- *Common promises:*
  - More predictive preclinical studies
  - Replacement of animal models
  - Patient models for personalized medicine
  - In-depth disease modelling
- *Common problems:*
  - Low maturity
  - Lack of complexity (architecture, cell types, biomechanics, biochemical environment)
  - Insufficient validation, reproducibility
  - Lack of physiologically relevant readouts

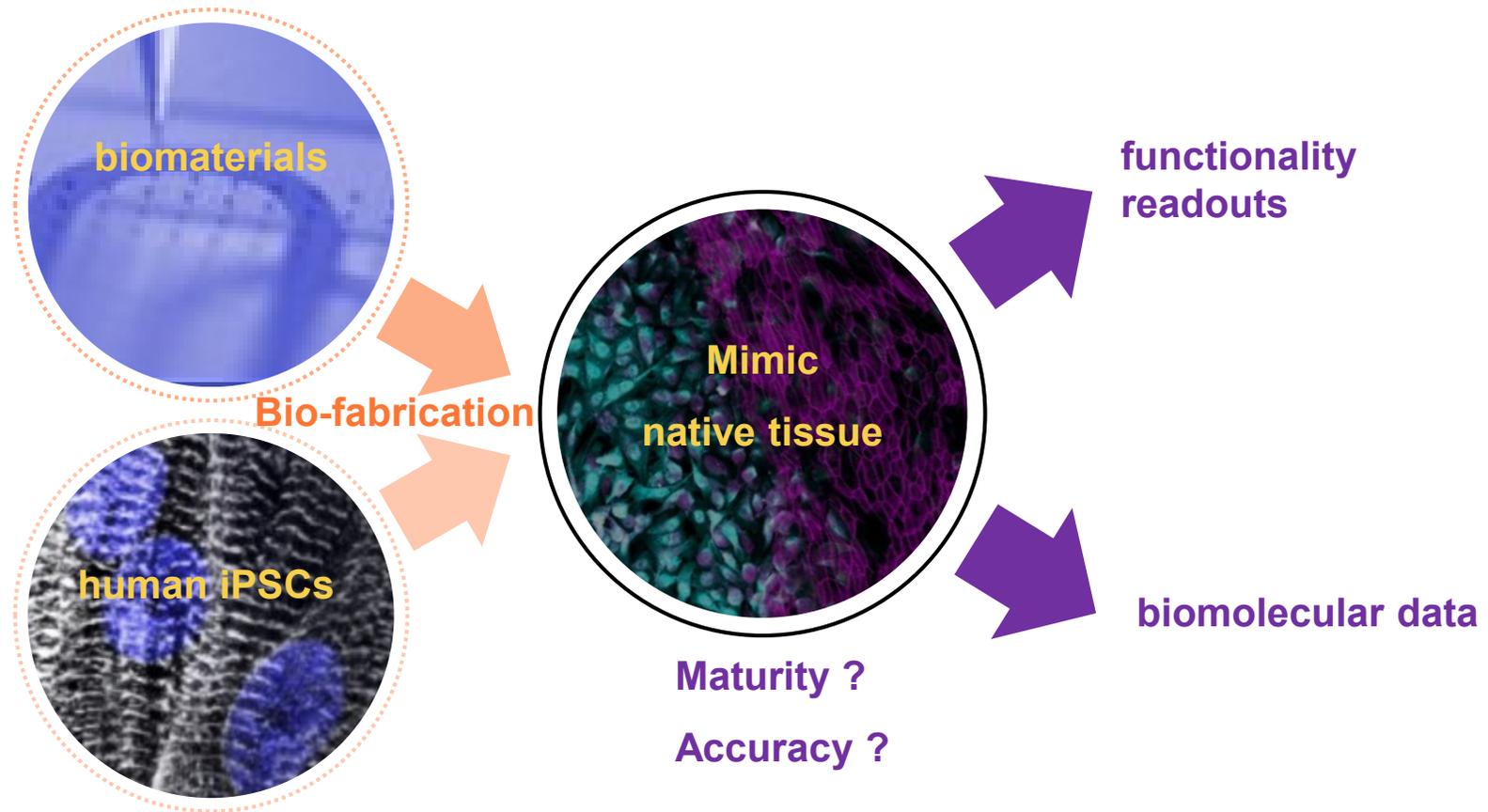


Human stem cell-derived tissues

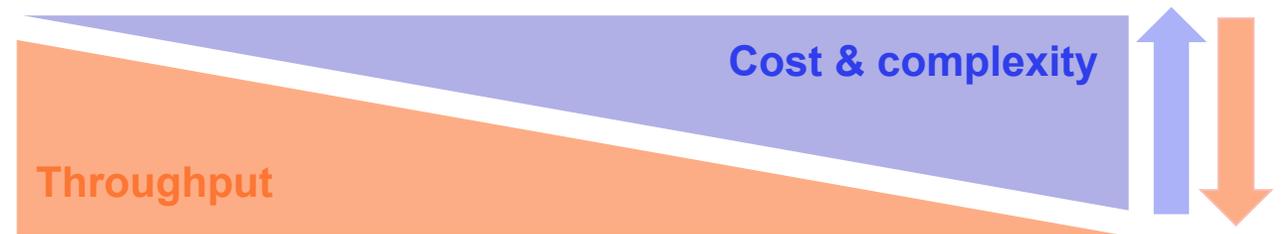
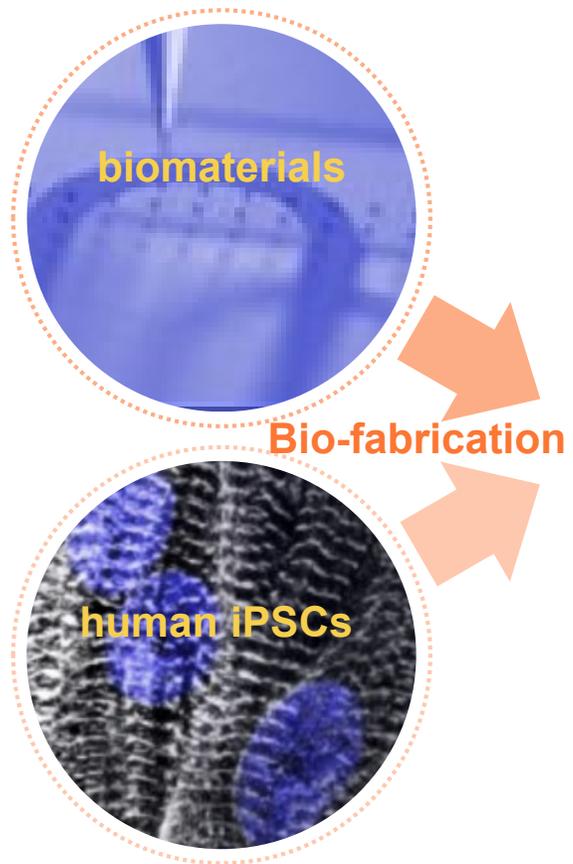
Lind *et al* Lab on a Chip 2017



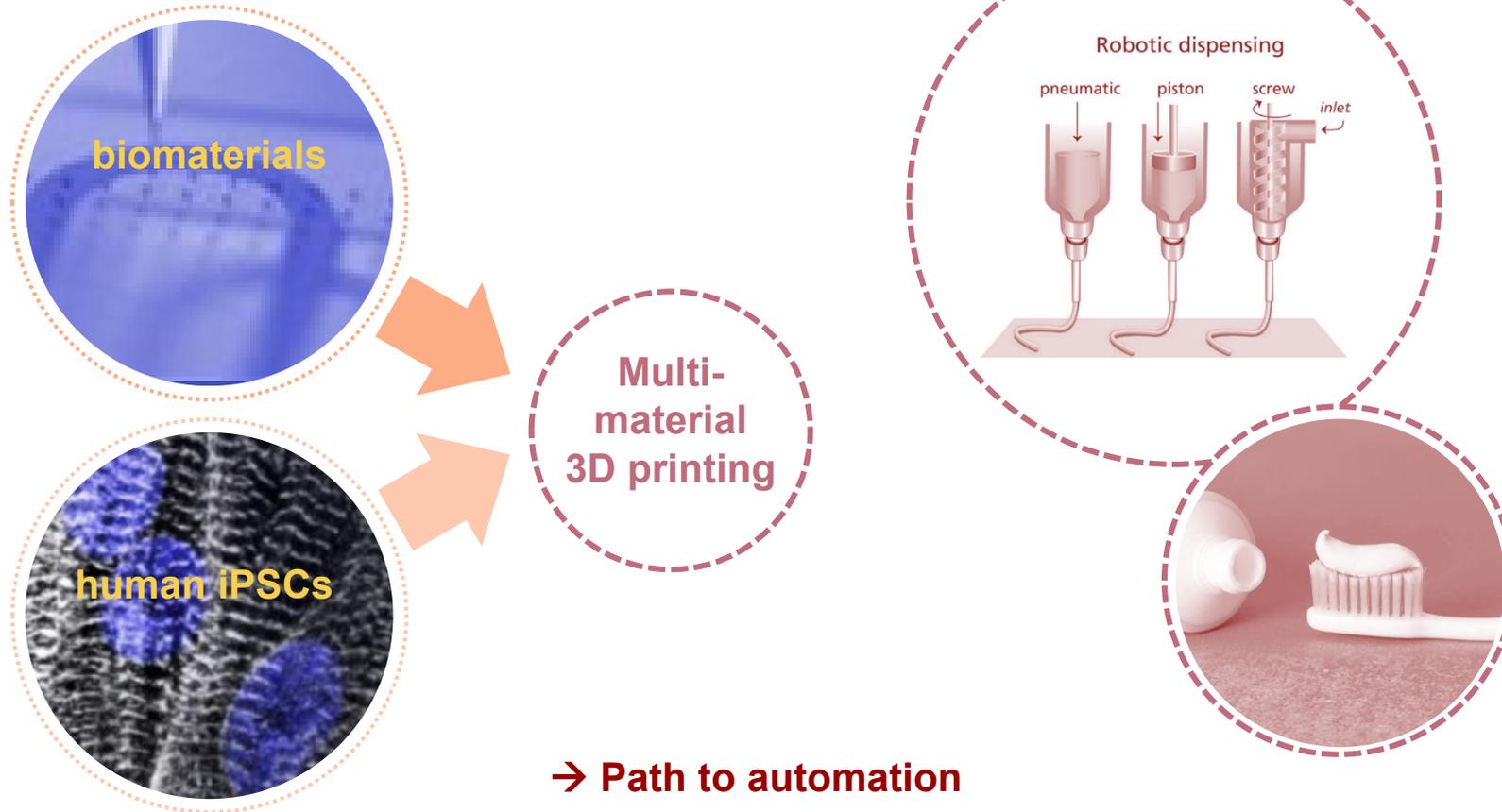
# Engineering challenges



# Engineering challenges



# Engineering challenges

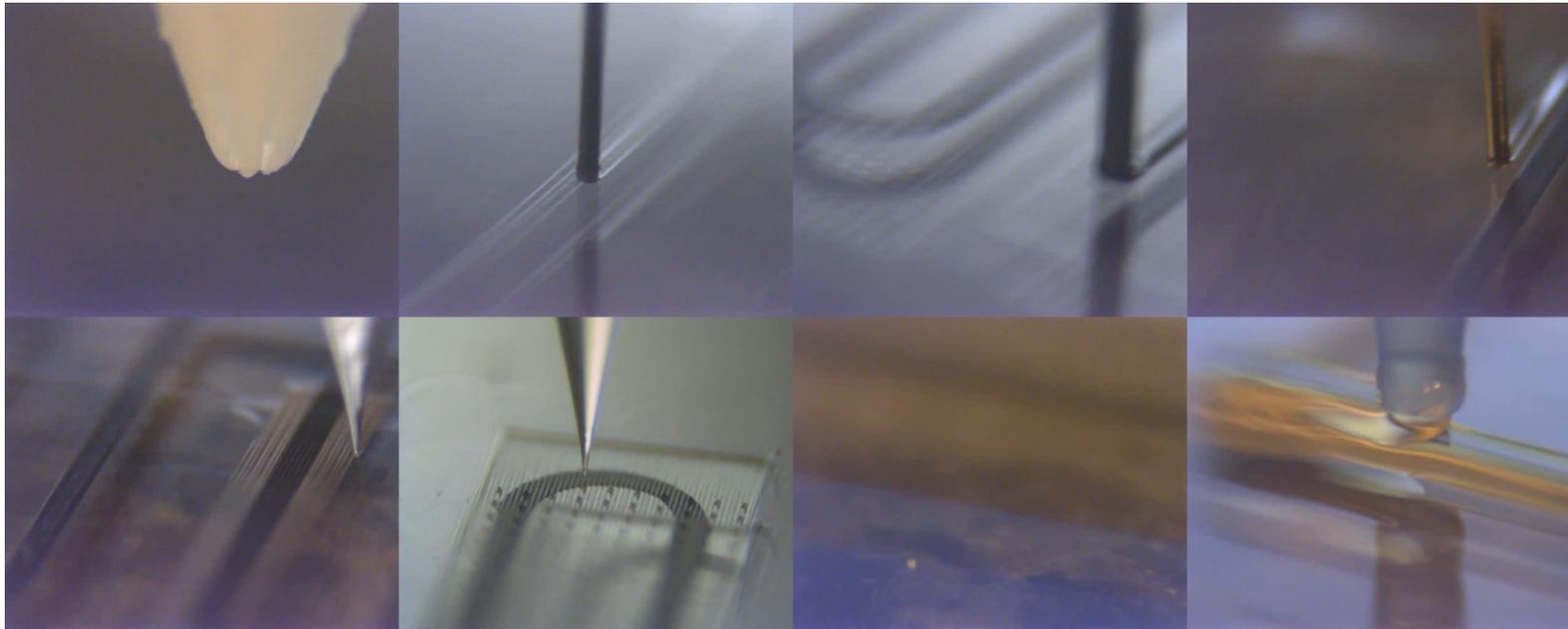


→ Path to automation

→ Customization across complexity space?

# Instrumented human cardiac tissue models

# Biofabrication using Multi-material 3D printing



# Cardiac Muscle

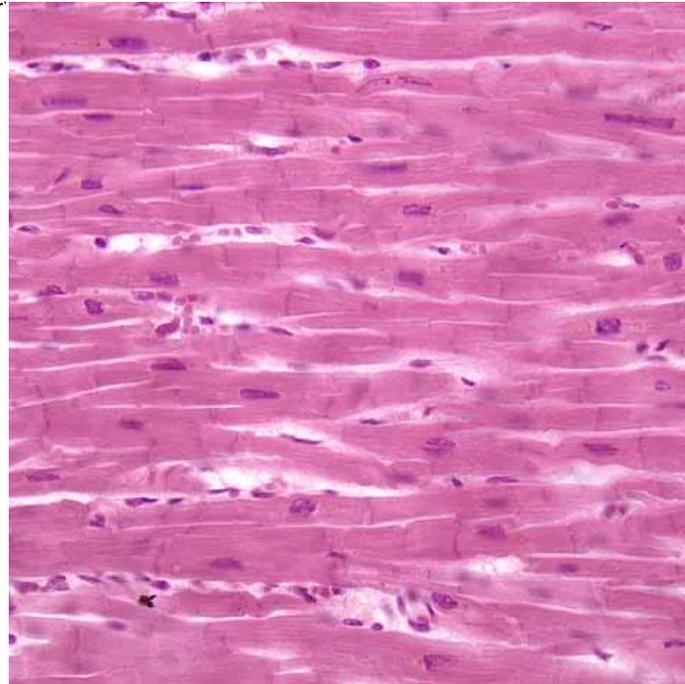
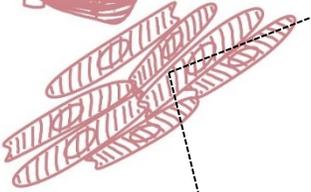
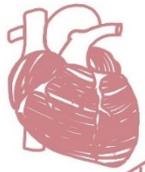
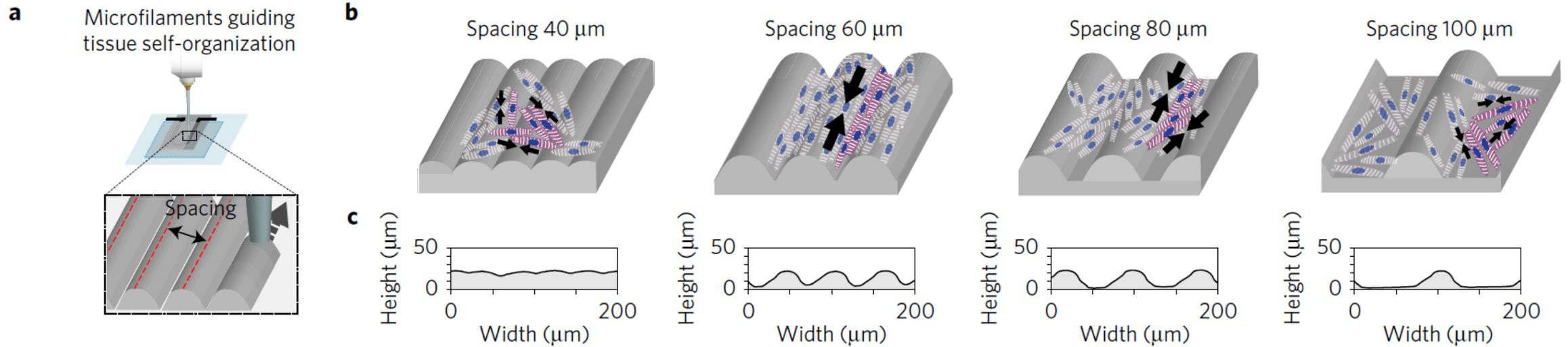


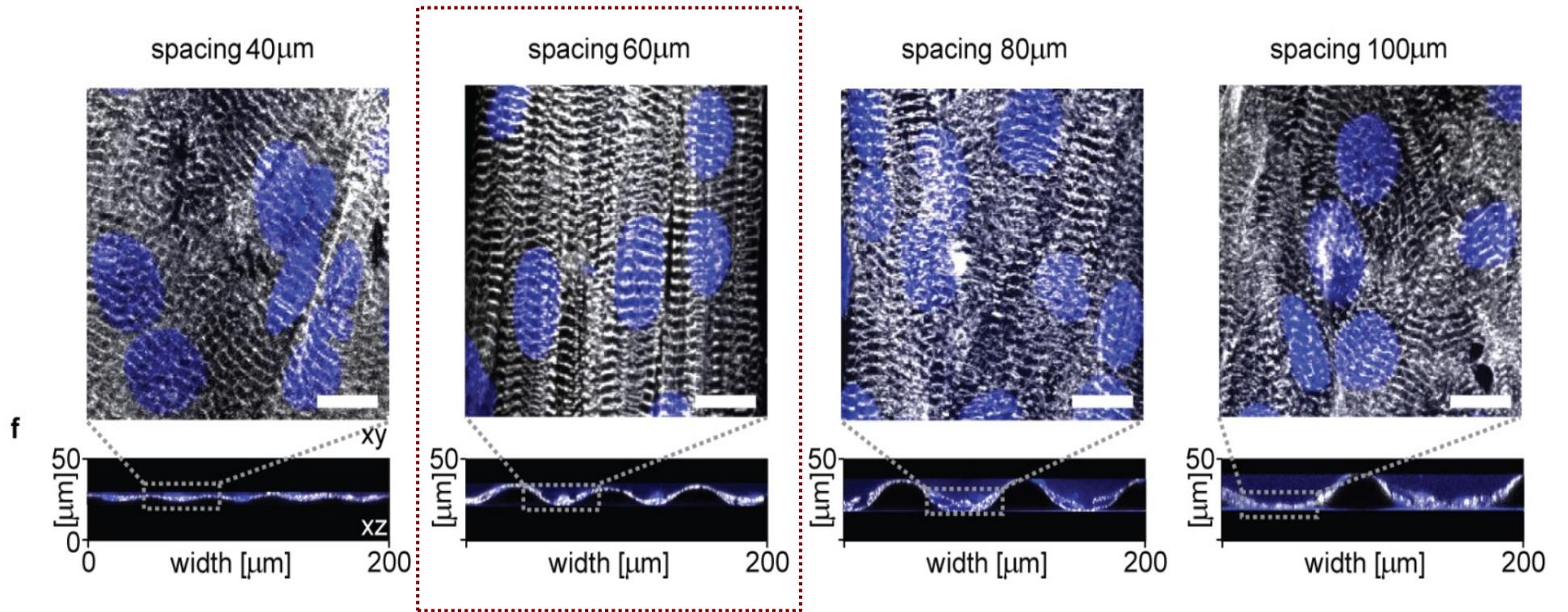
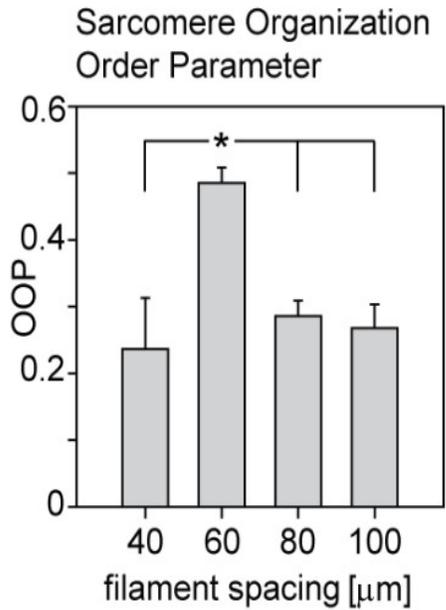
Image courtesy of google

- **anisotropic and laminar**
- **myocyte sheets ~4 cells thick**
- **highly vascularized**

# Printed Microstructures Guide Muscle Architecture



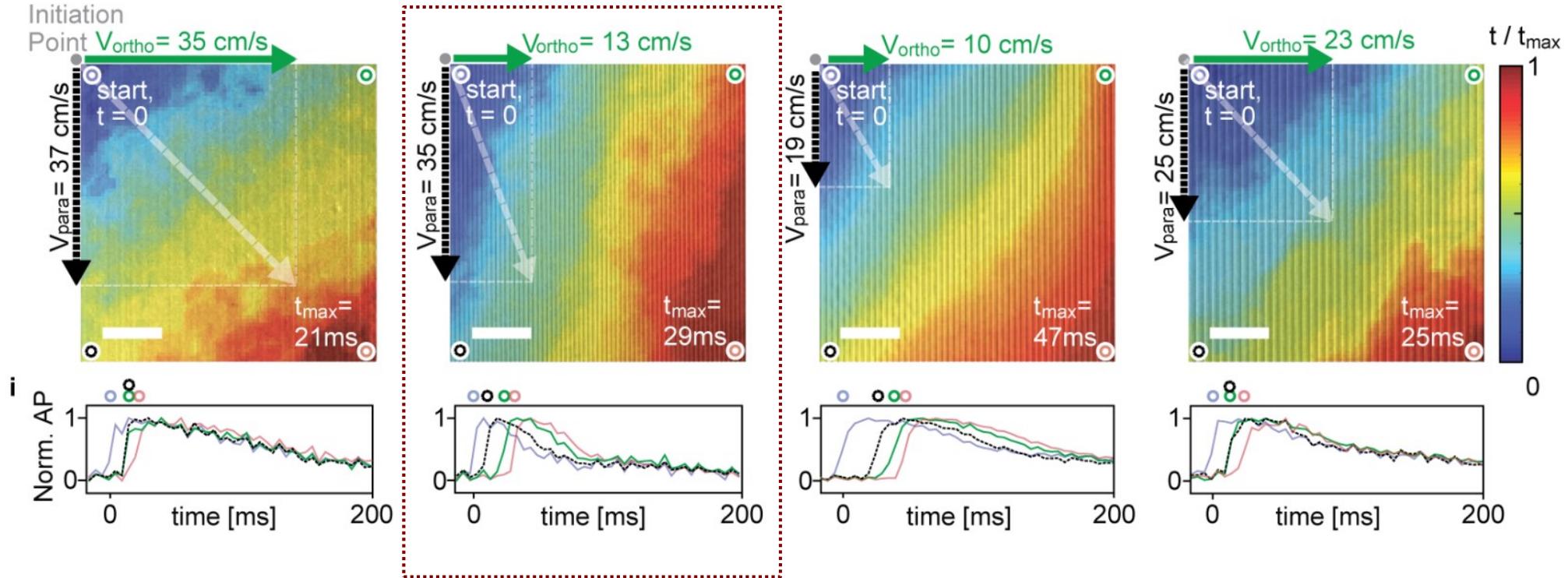
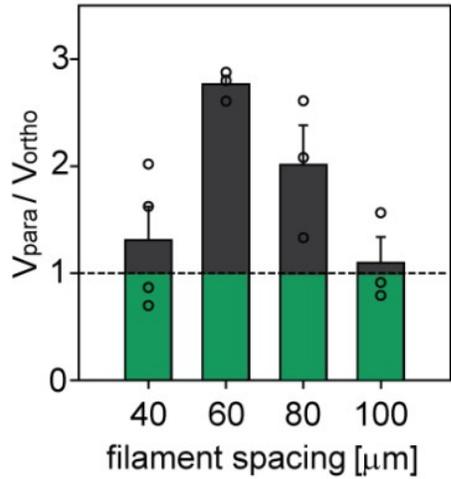
# Mimicking native structure



OOP of 0.5 indicative of highly aligned sarcomeres

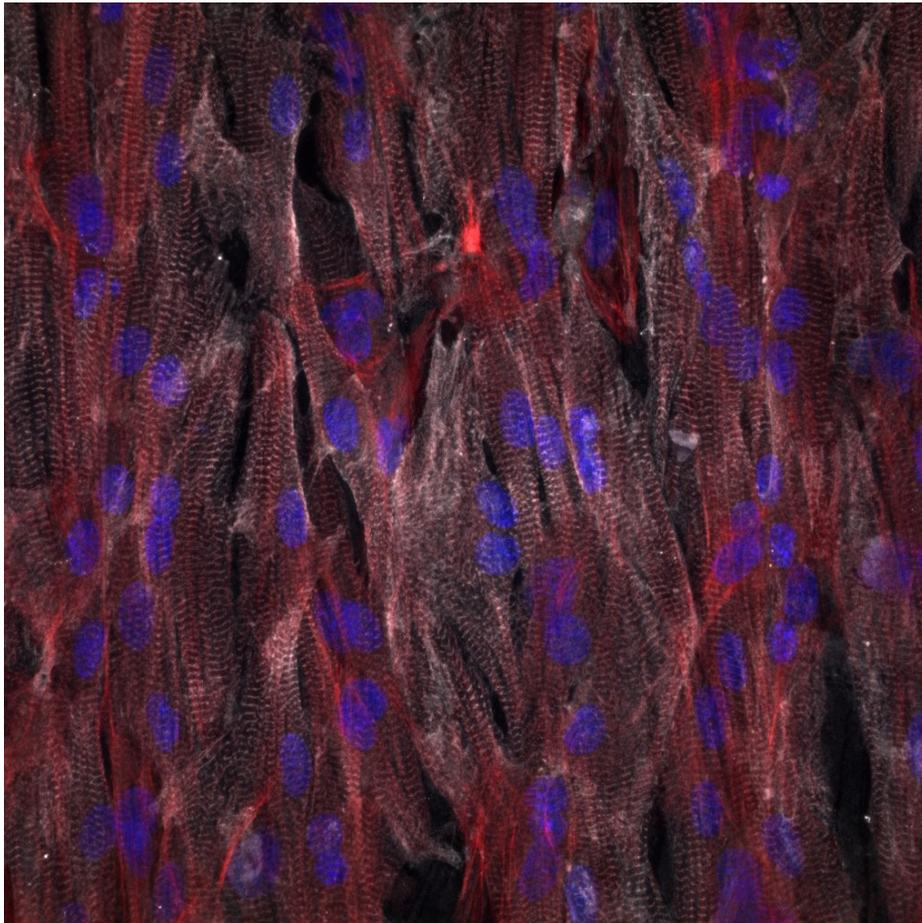
# Mimicking native electrophysiology

Action Potential Propagation Velocity

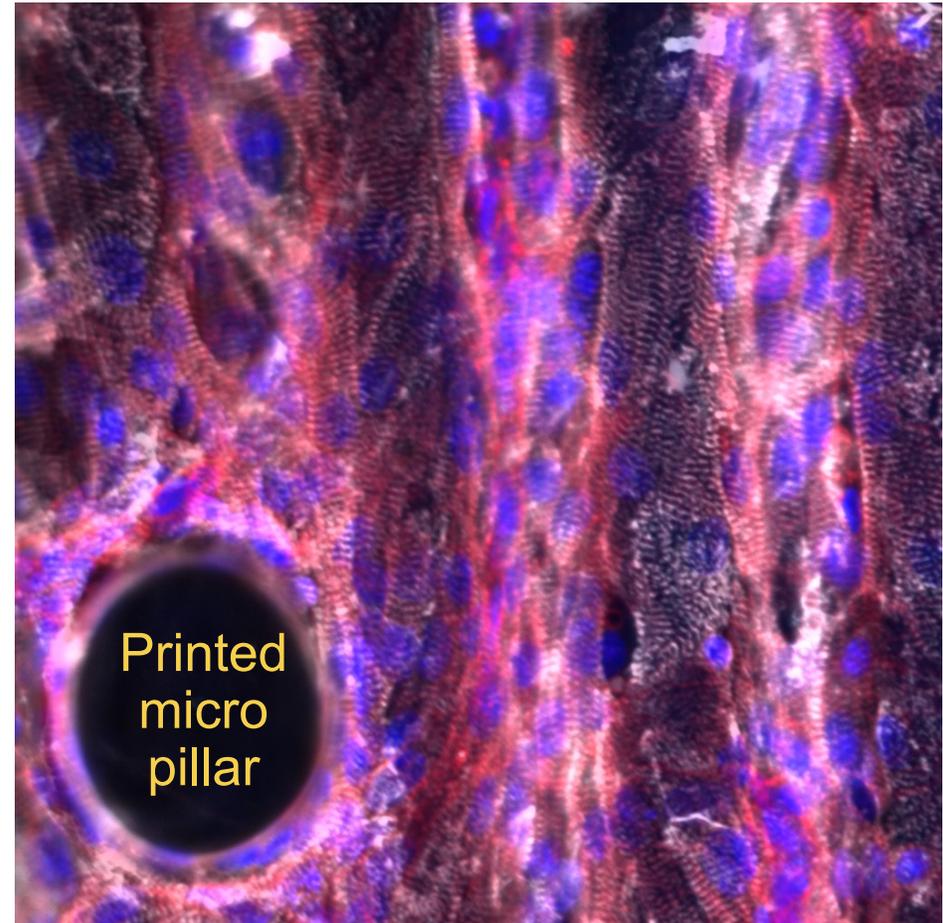


AP propagation speed ratio 2.1 matches native ventricular rat tissue sheets

# Adjustable tissue thickness

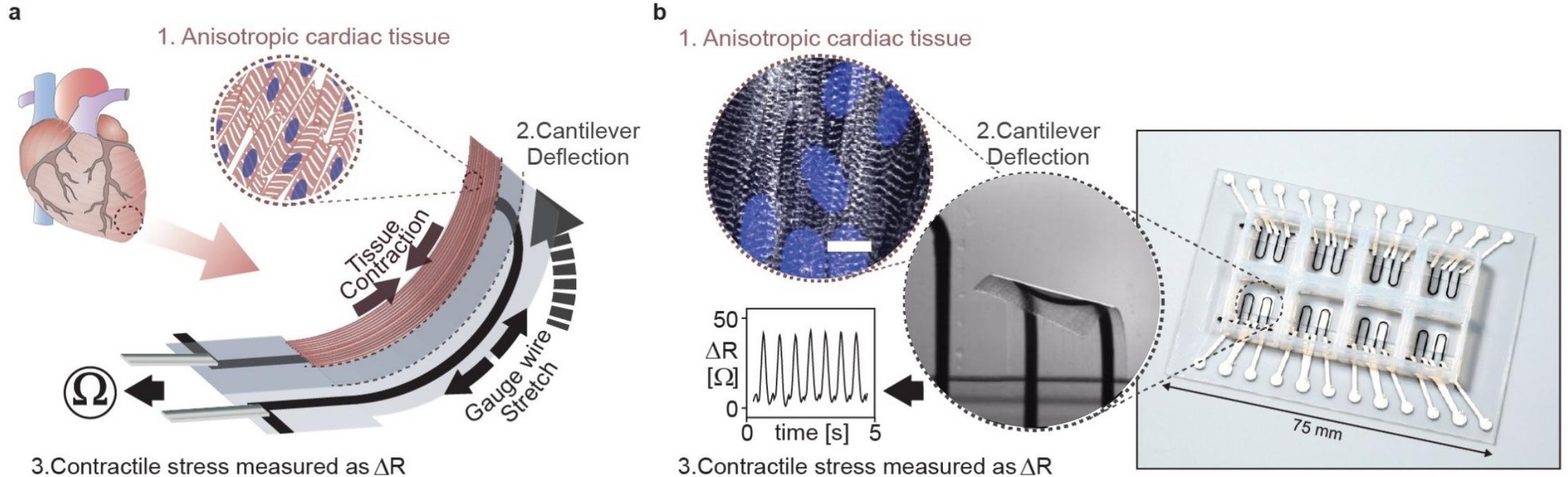


Monolayer CM tissue



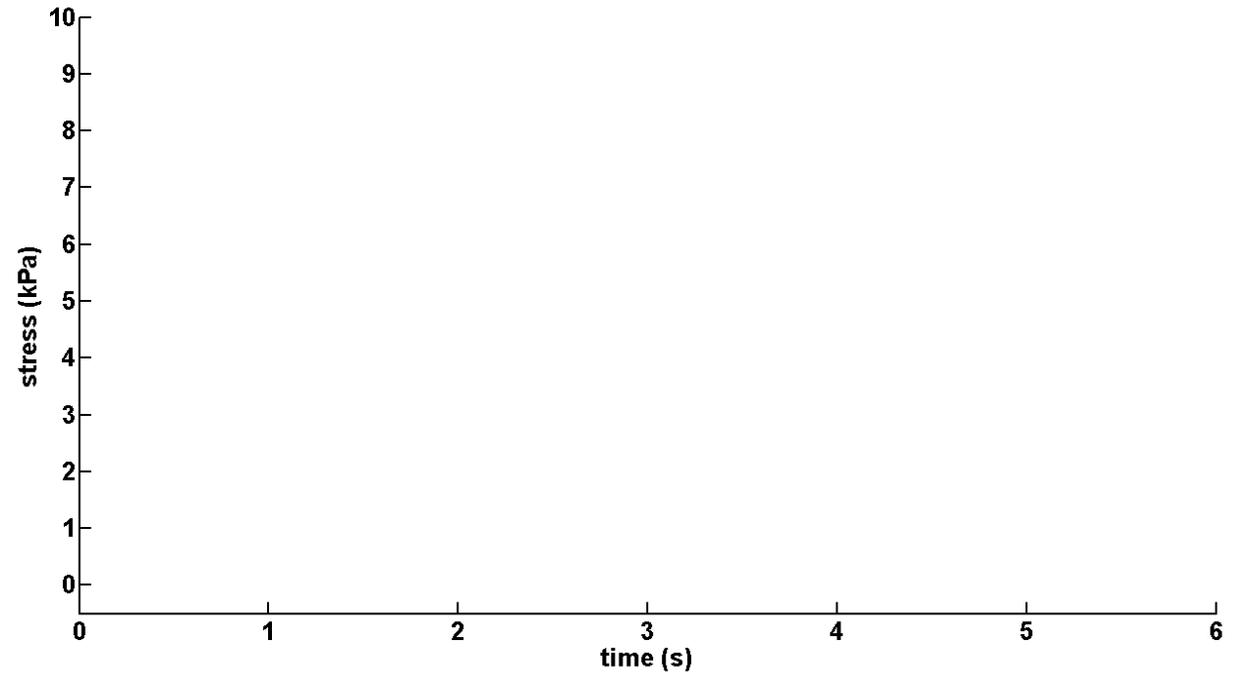
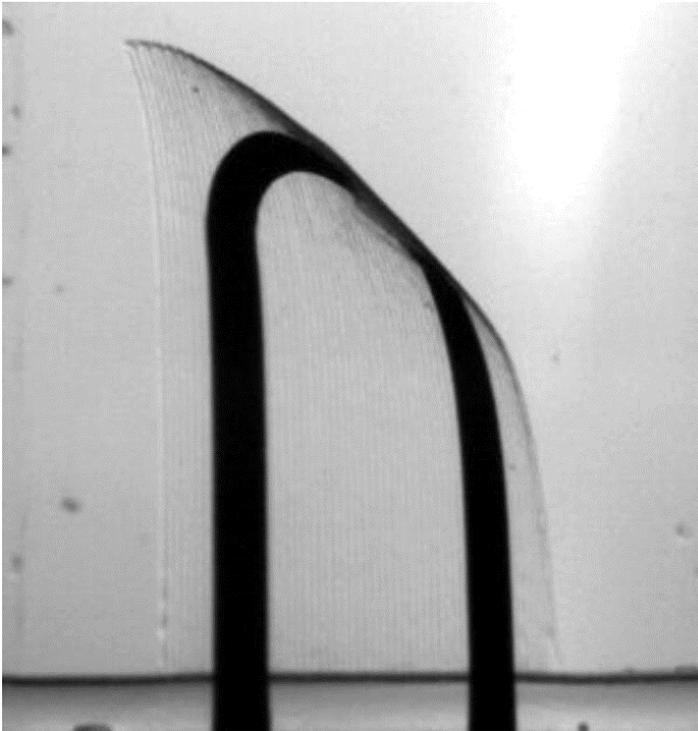
4-layer CM tissue (pillar anchor)

# 3D printed instrumented Heart on a Chip



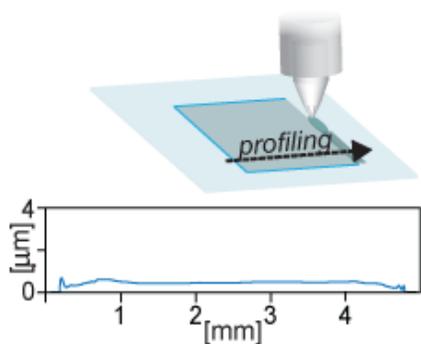
Adapted from JU Lind *Nature Materials* 2017

# 3D printed instrumented Heart on a Chip

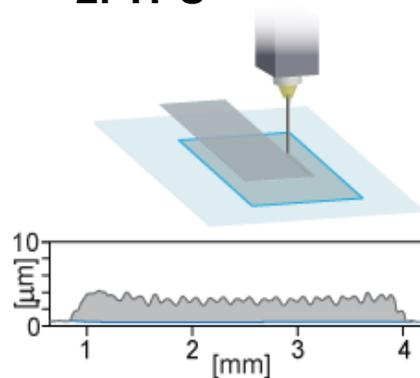


# Cantilever layered prints

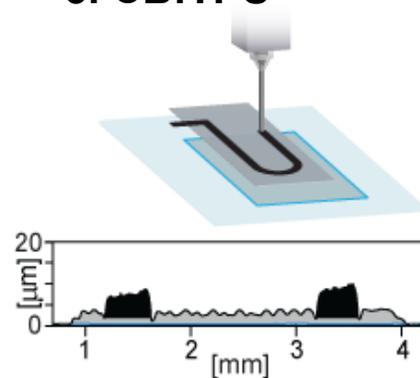
1. Dextran



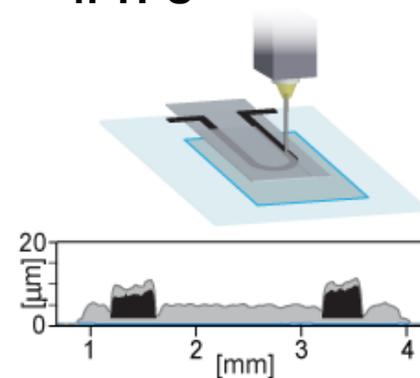
2. TPU



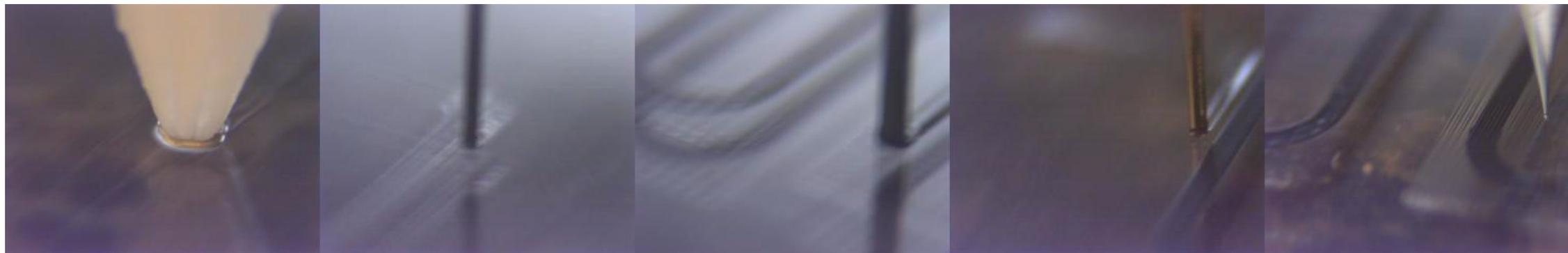
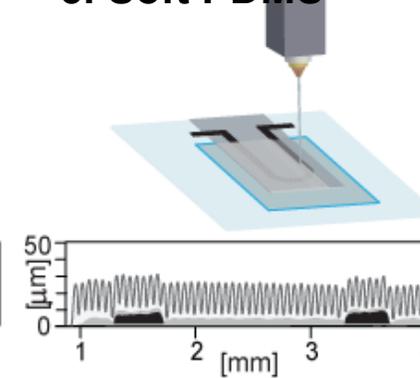
3. CB:TPU



4. TPU

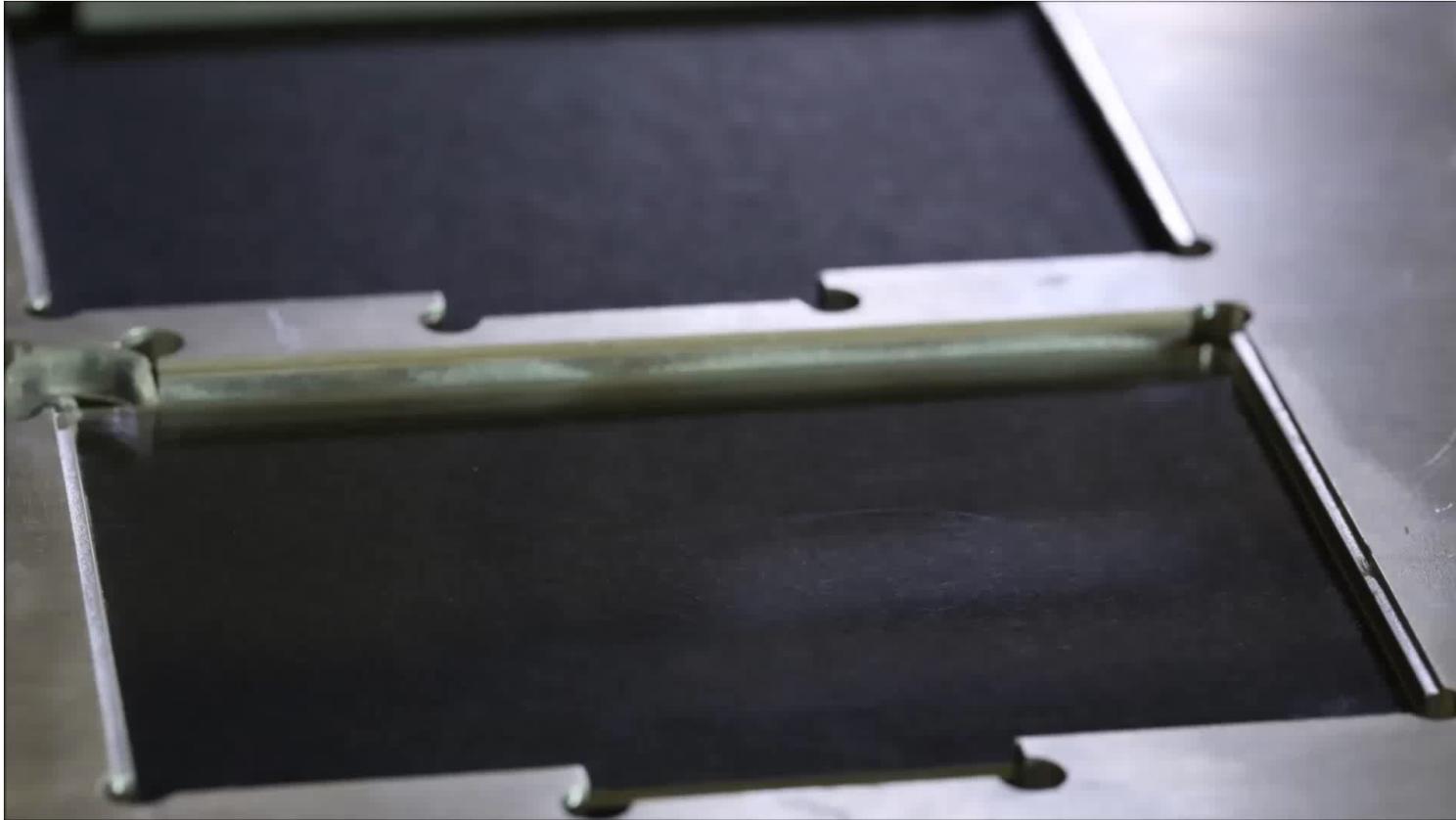


5. Soft PDMS

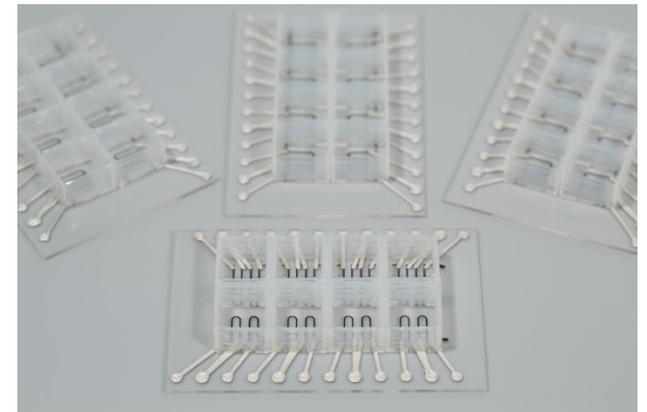


Adapted from JU Lind et al *Nature Materials* 2017

# Multimaterial printing of Heart Chips

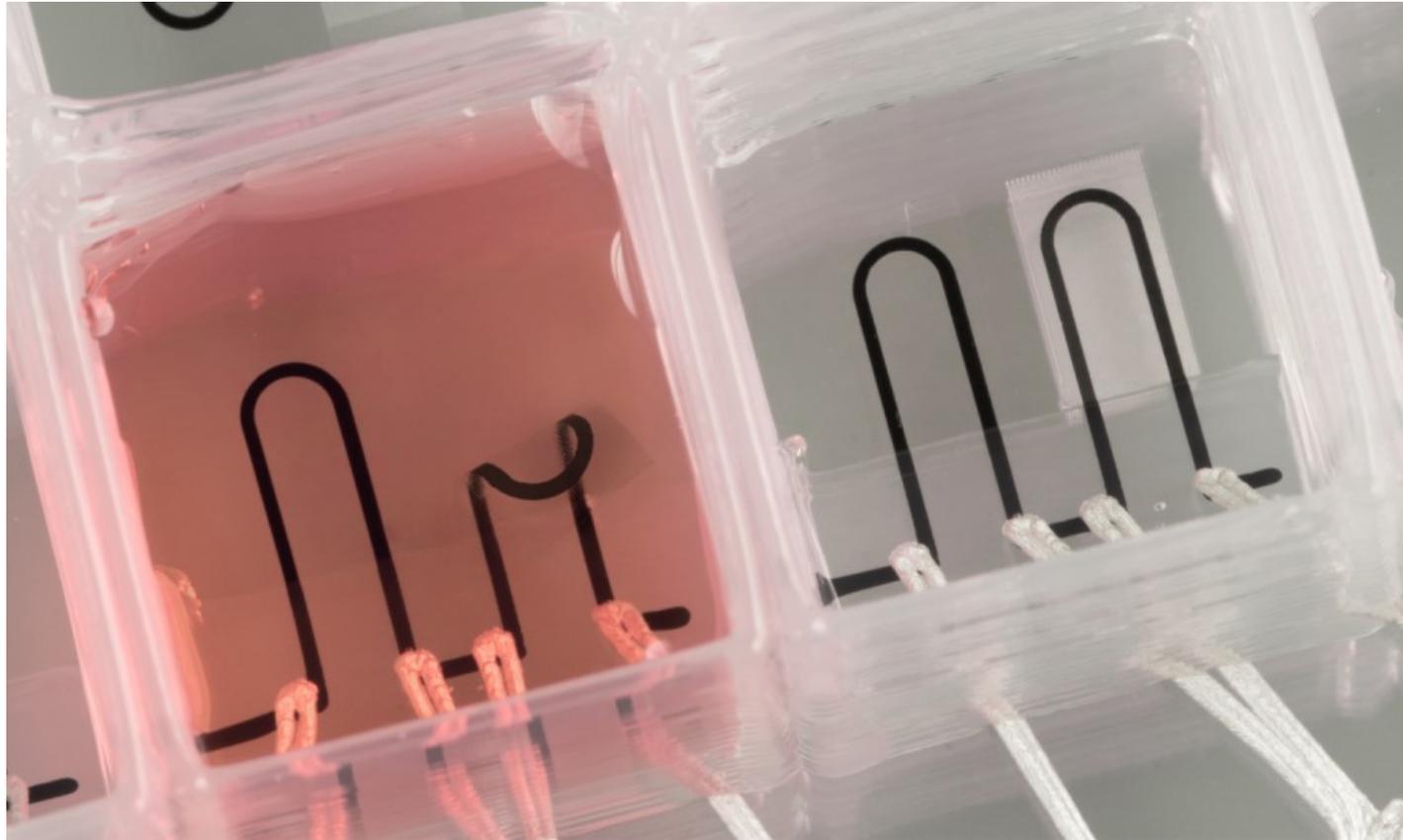


160x

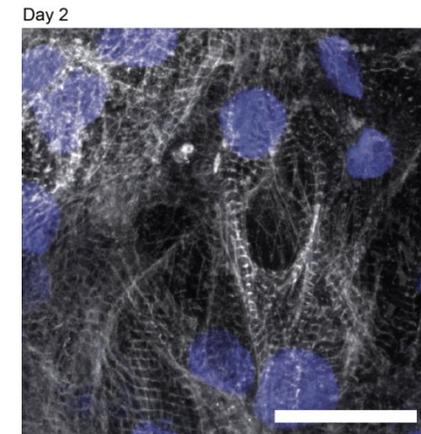
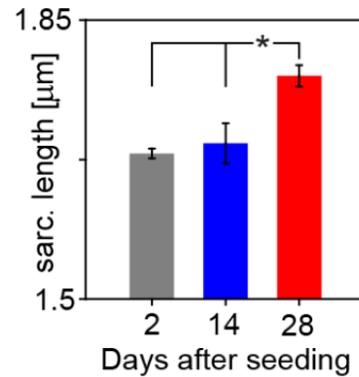
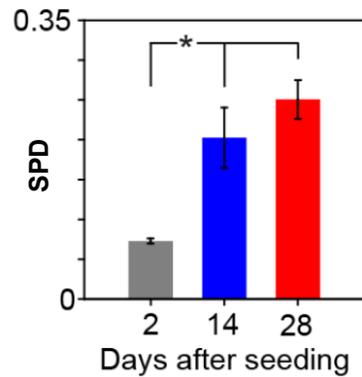
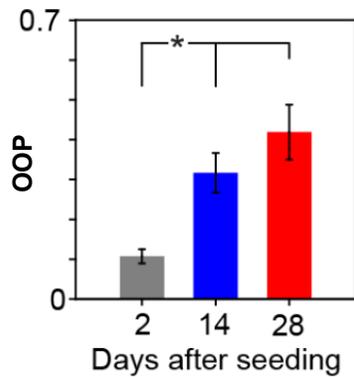
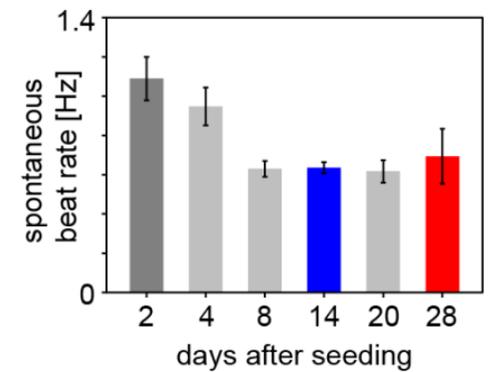
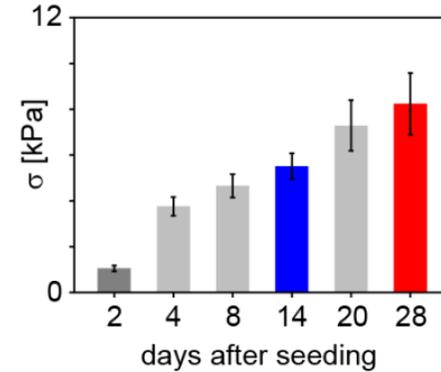
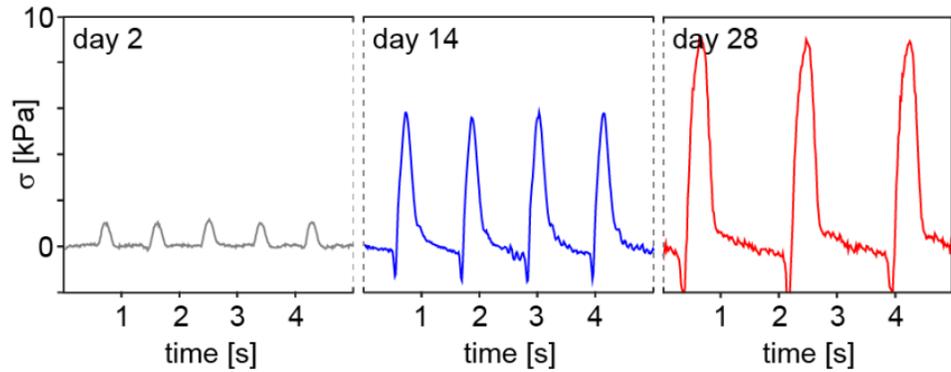


Adapted from JU Lind et al *Nature Materials* 2017

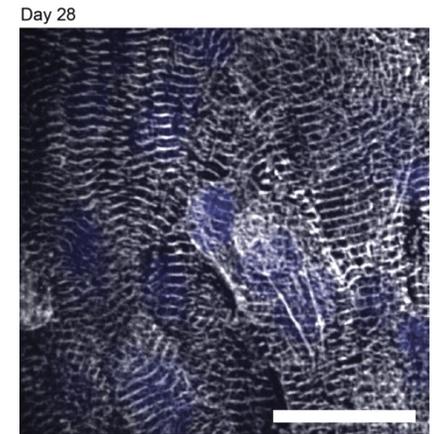
# Instrumented heart chip applications



# Long term maturation of Human iPS-CM Tissue

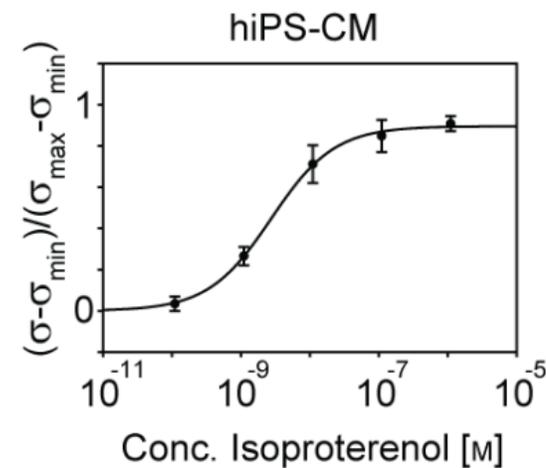
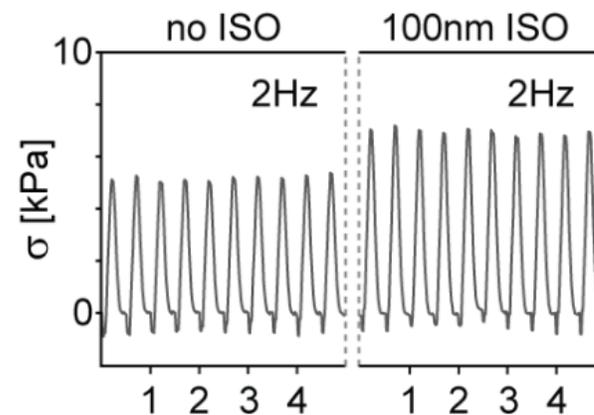
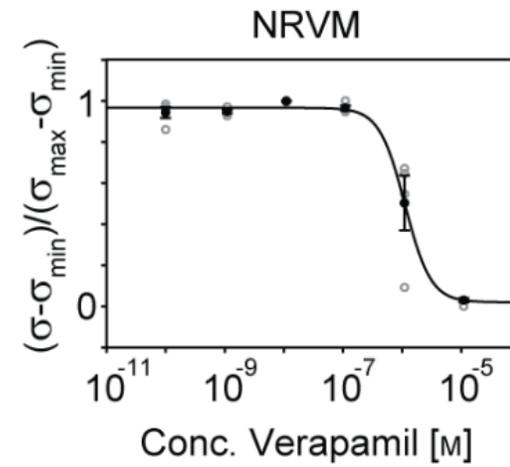
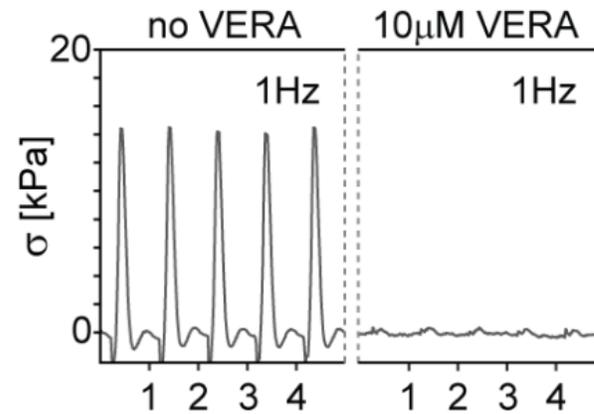


OOP = 0.11

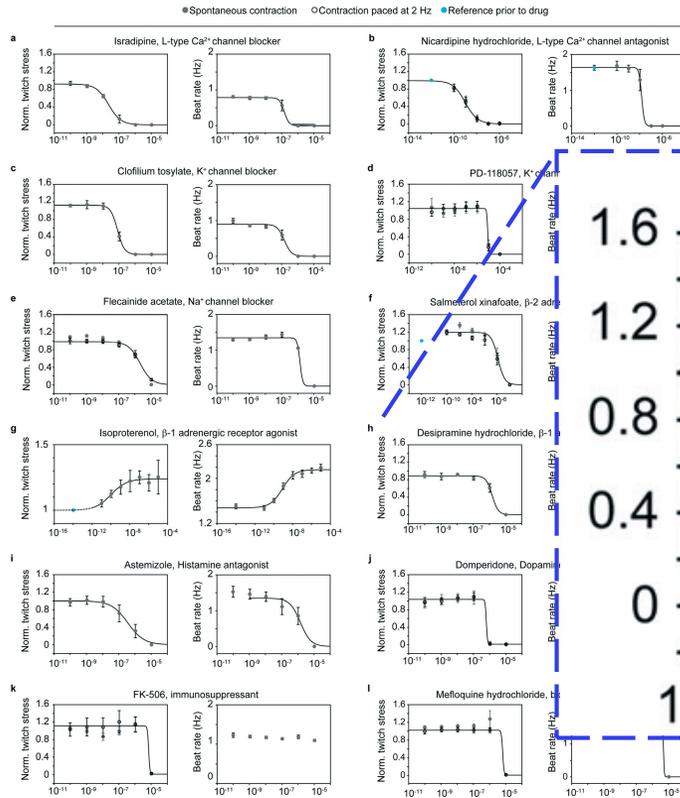
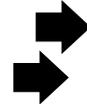
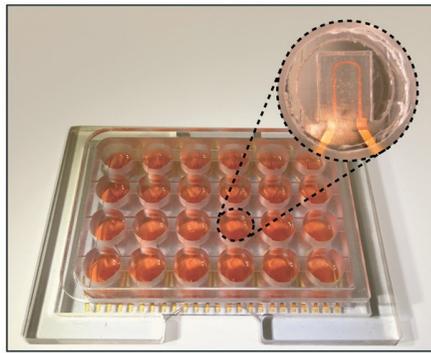


OOP = 0.42

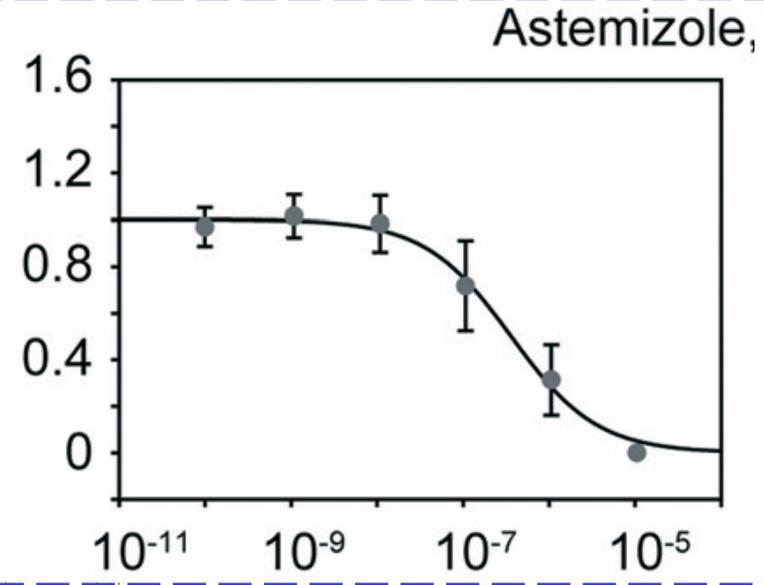
# Example drug tests



# Drug toxicity tests



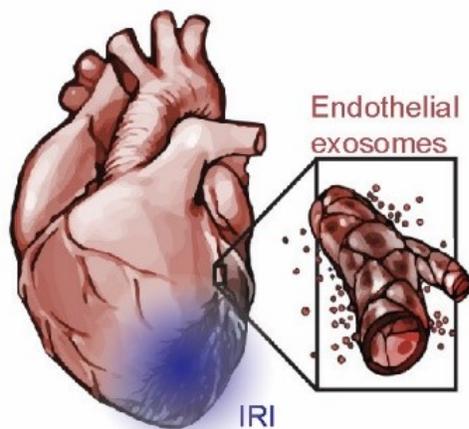
**Astemizole Antihistamine (1977)**  
**HERG block side-effects**  
**Withdrawn 1999**



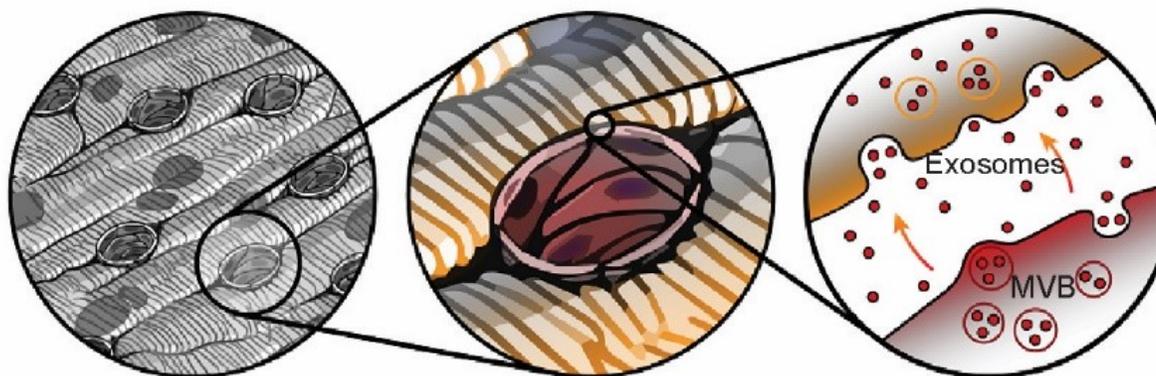
Adapted from JU Lind et al *Lab Chip* 2017

# Simulated ischemic heart failure

**A** In vivo  
Ischemic reperfusion injury

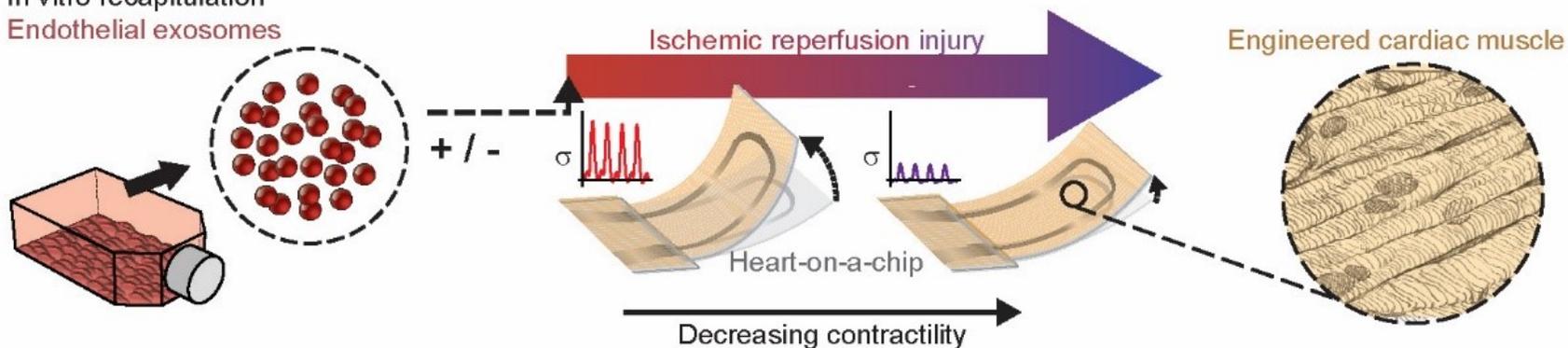


Cardiac muscle: Cardiomyocyte / Receptient Cell



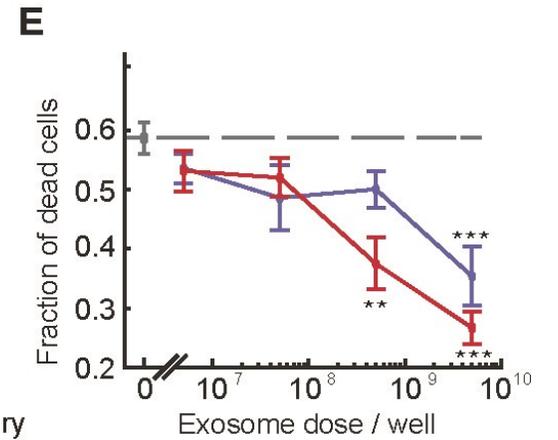
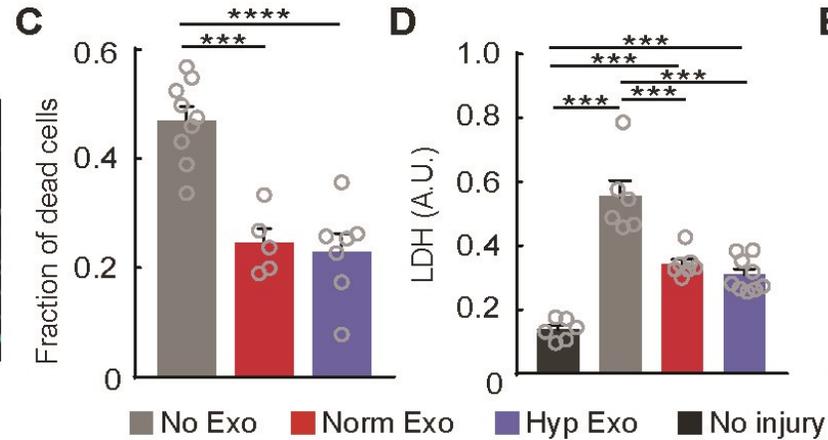
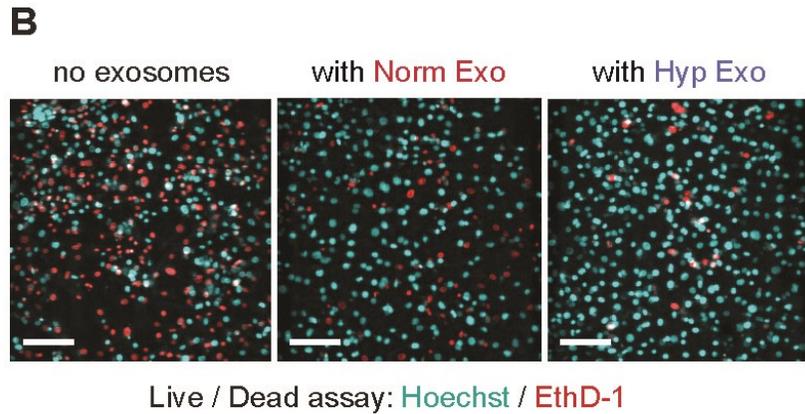
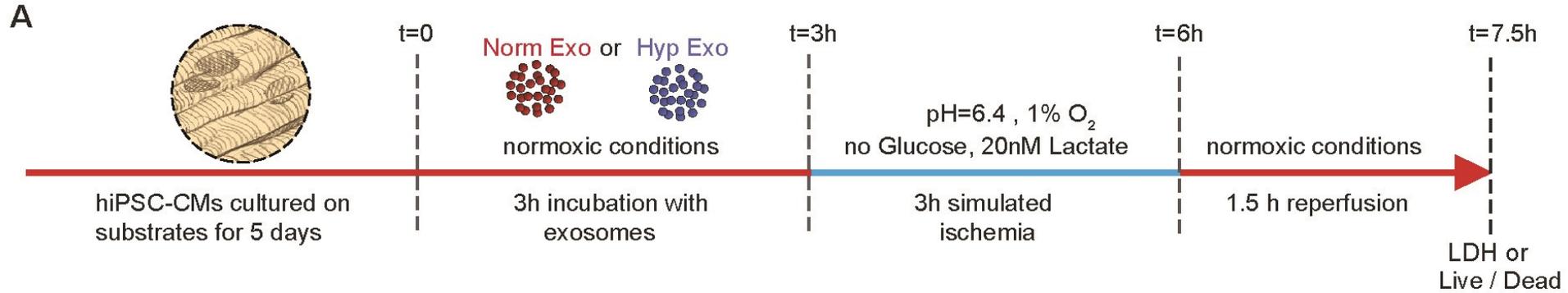
Vasculature: Endothelial Cell / Donor Cell

**B** In vitro recapitulation  
Endothelial exosomes



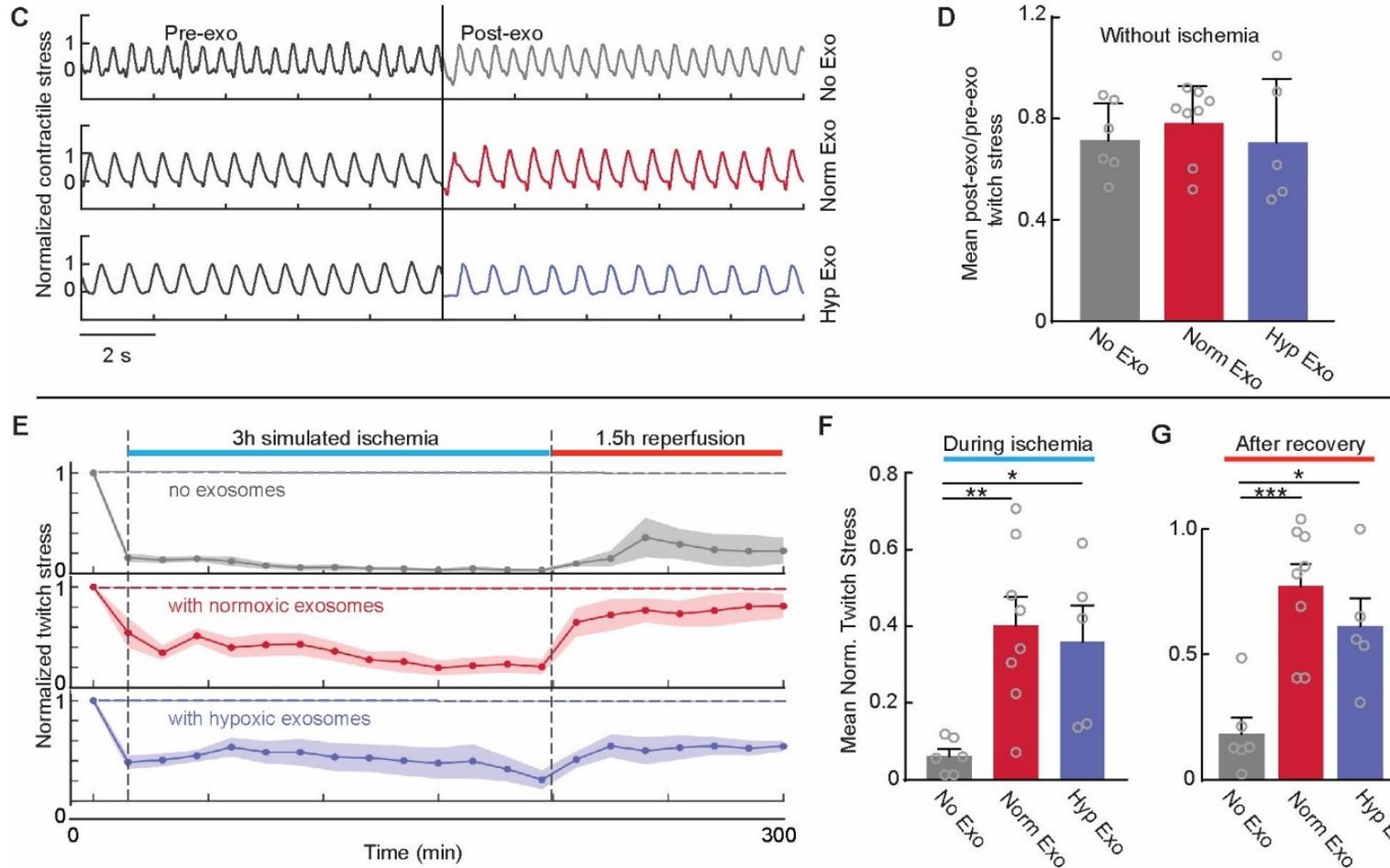
Adapted from M Yadid et al *Science Translational Medicine* 2020

# Simulated ischemic heart failure



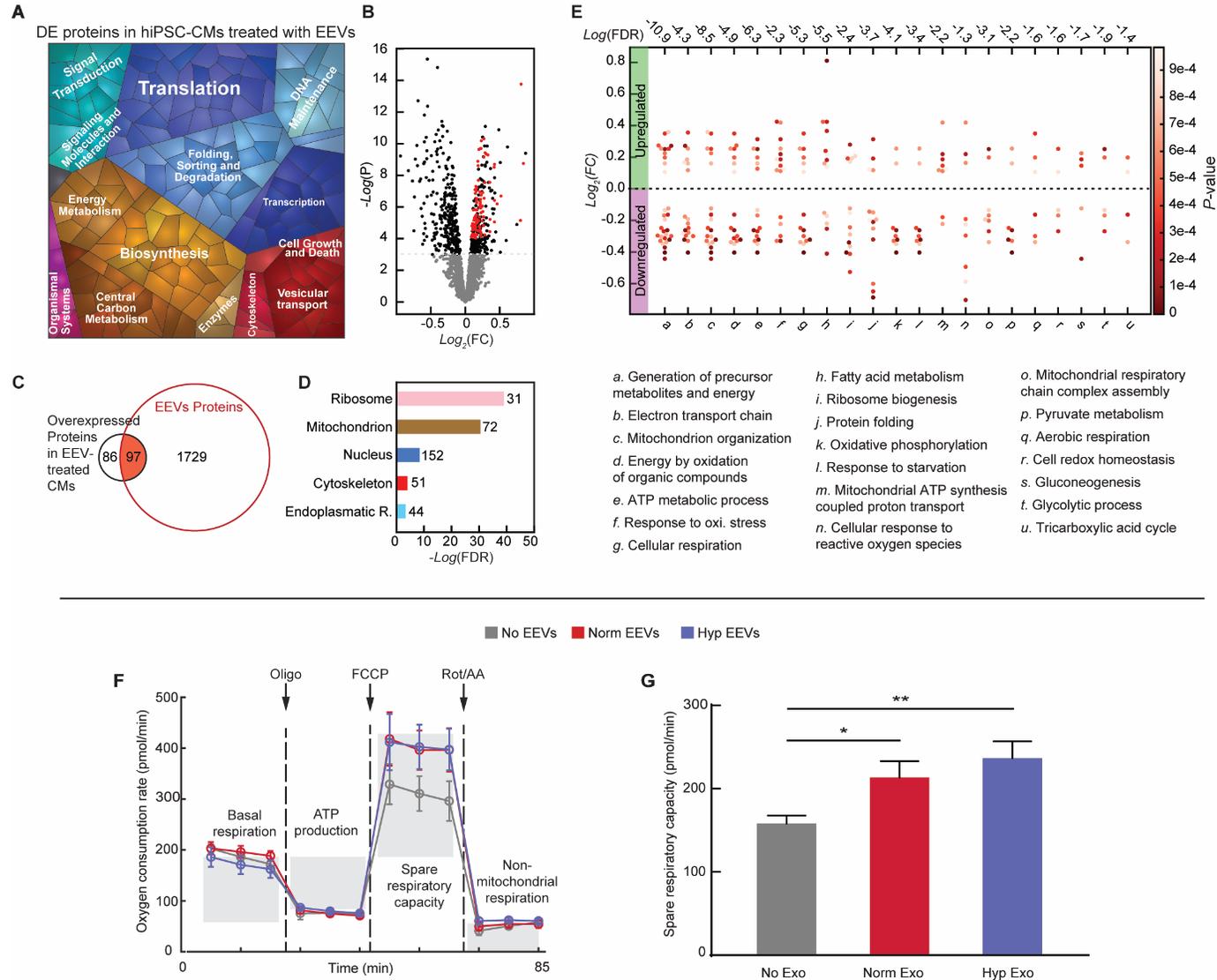
Adapted from M Yadid et al *Science Translational Medicine* 2020

# Simulated ischemic heart failure



Adapted from M Yadid et al *Science Translational Medicine* 2020

# EEVs increase CM respiratory capacity

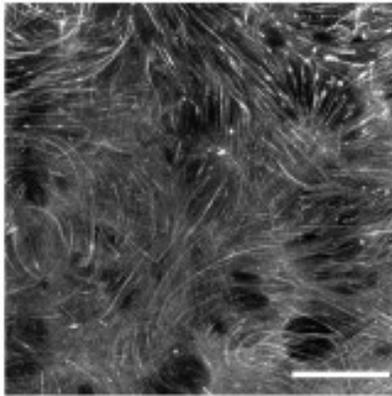


Adapted from M Yadid et al *Science Translational Medicine* 2020

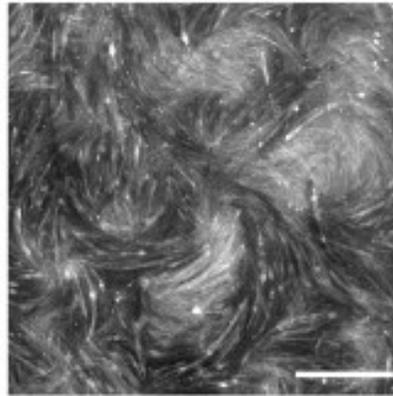
# Cell-instructive biomaterials

# Spontaneous architecture on ultra soft gelatin

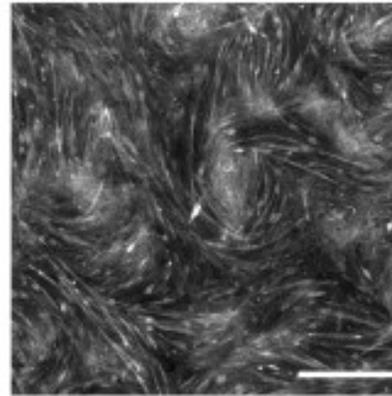
TC-PS



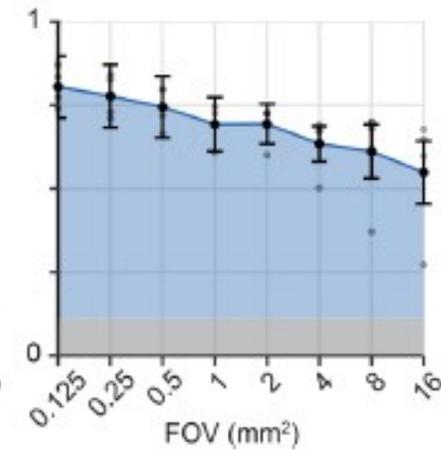
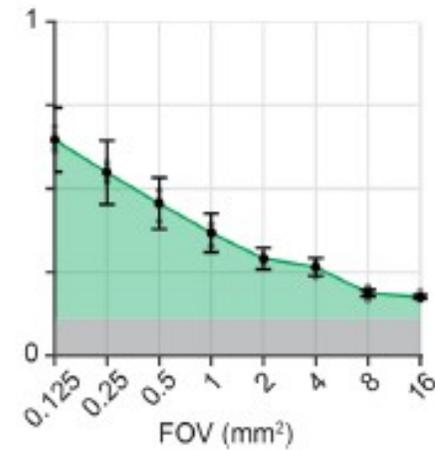
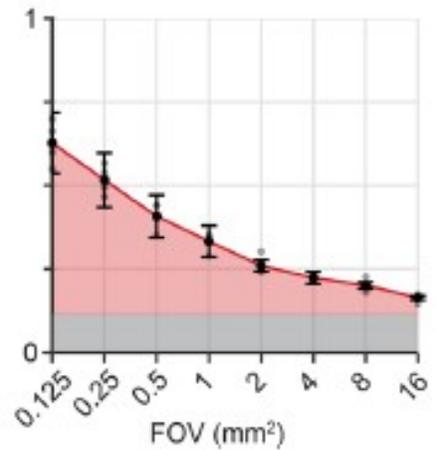
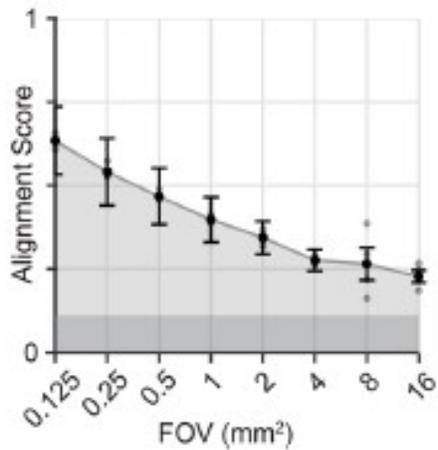
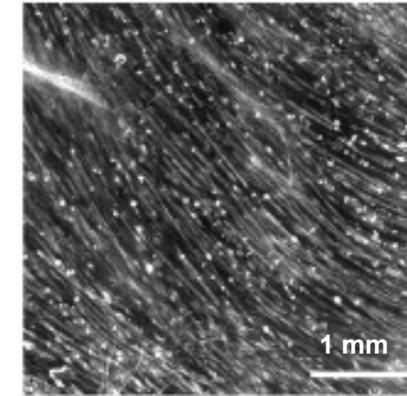
10% Gelatin, 10U/ml TG



5% Gelatin, 10U/ml TG

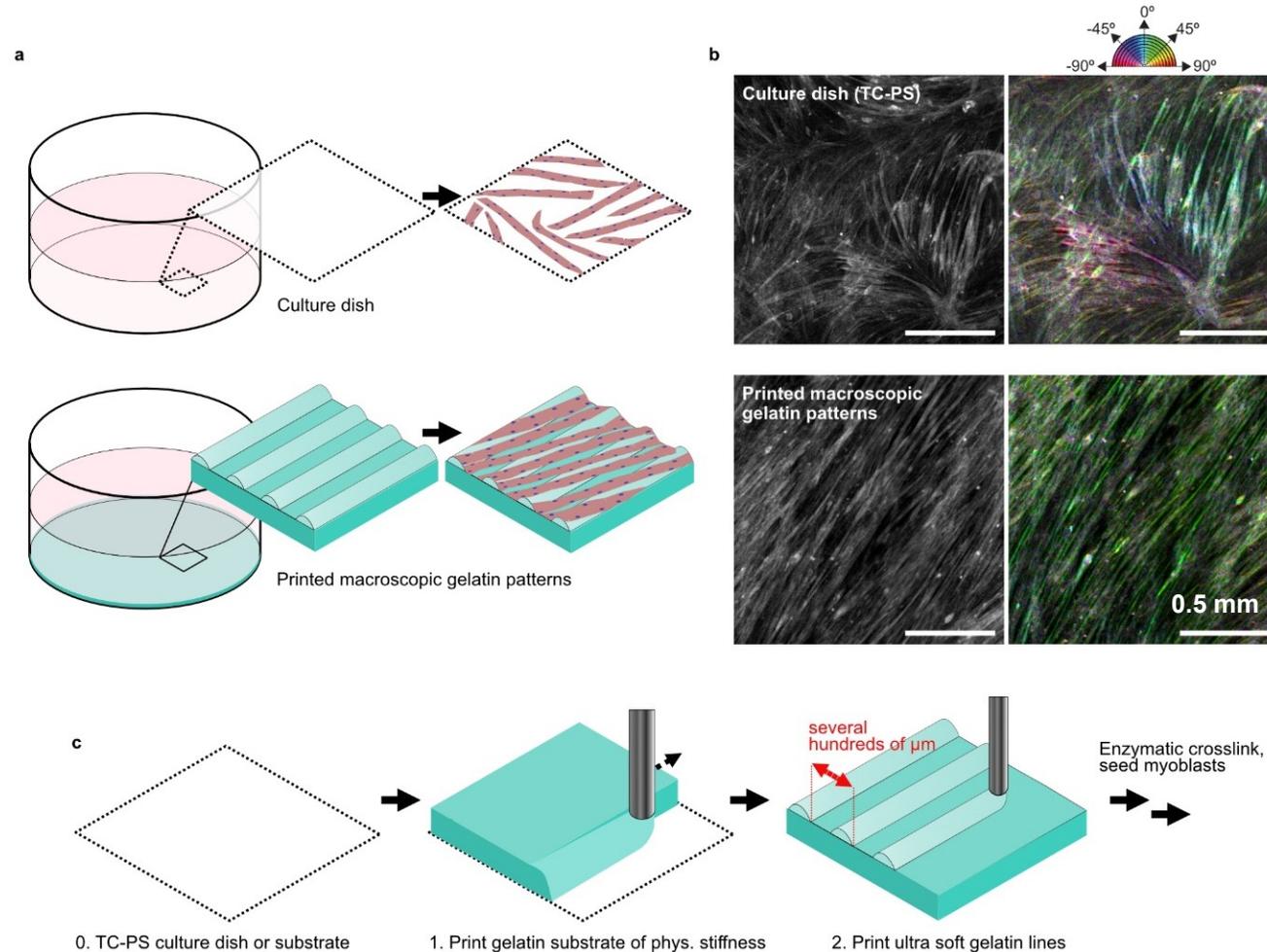


2.5% Gelatin, 10U/ml TG



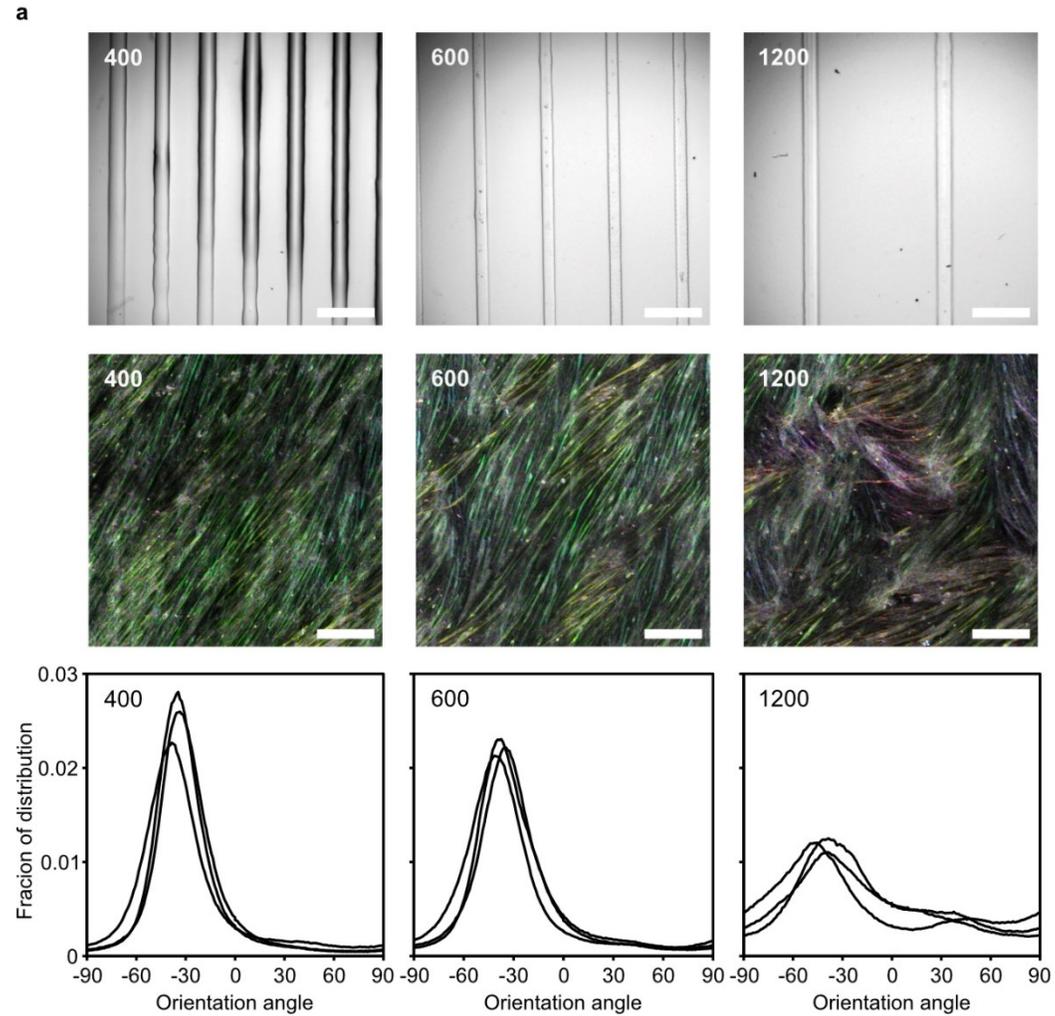
Adapted from JH Jensen et al *Scientific Reports* 2020

# Minimally Complex Printed Alignment Structures



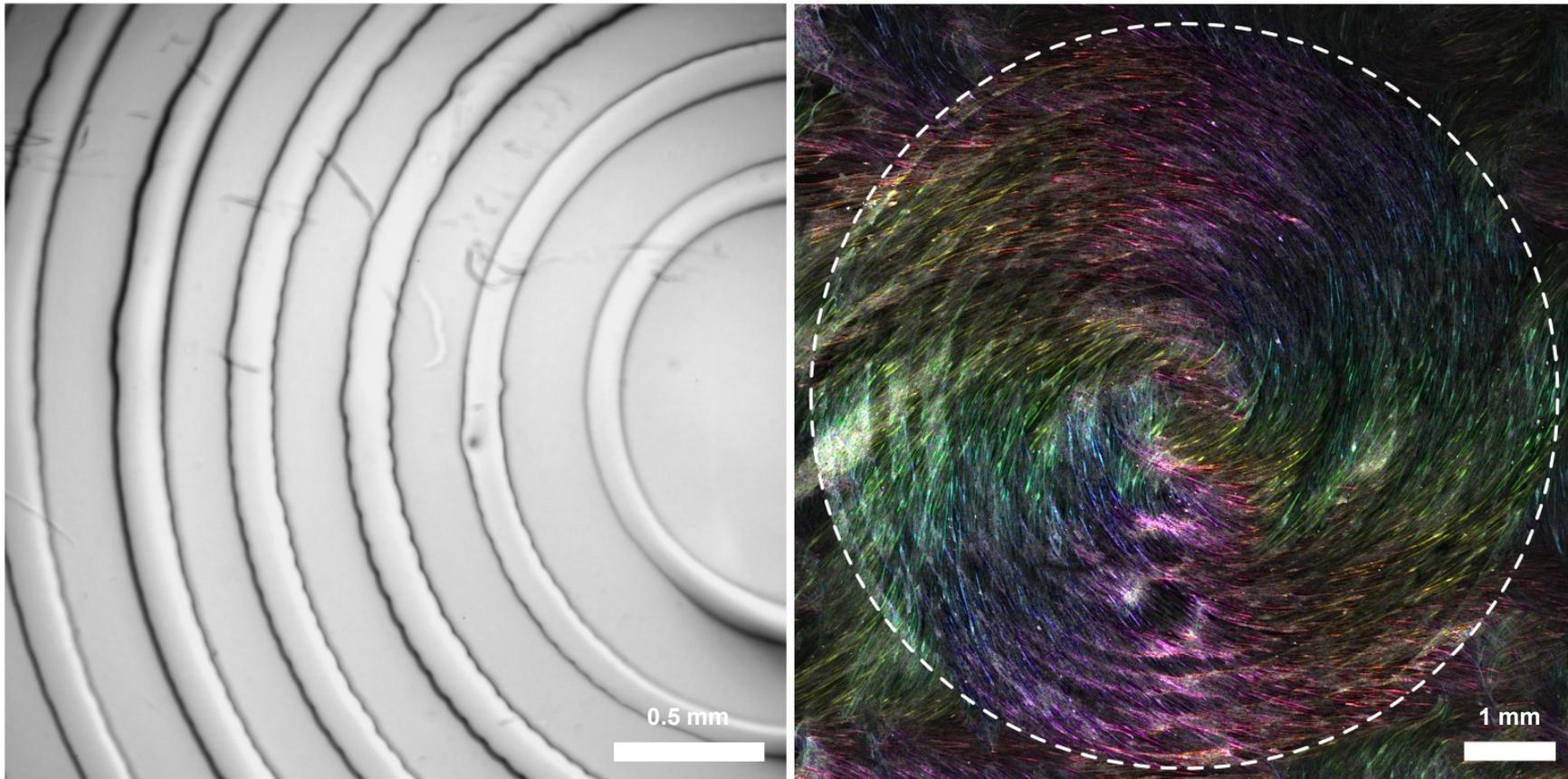
Adapted from SD Cakal et al *Biomedical materials* 2022

# Minimally Complex Printed Alignment Structures



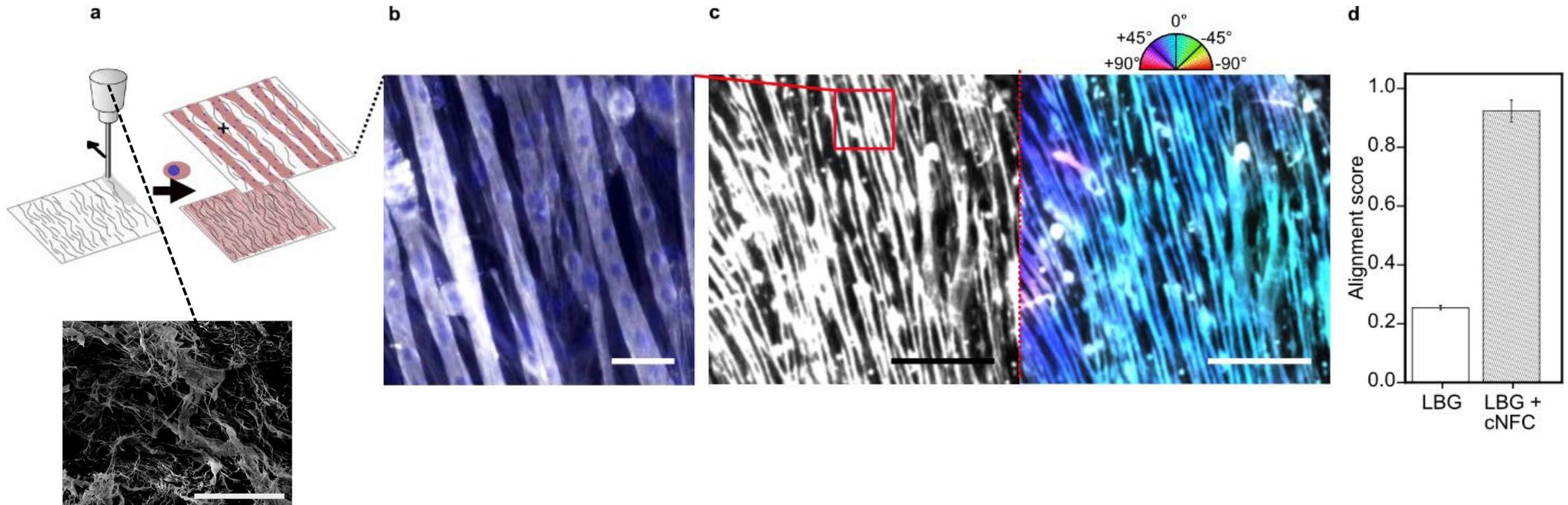
Adapted from SD Cakal et al *Biomedical materials* 2022

# Minimally Complex Printed Alignment Structures



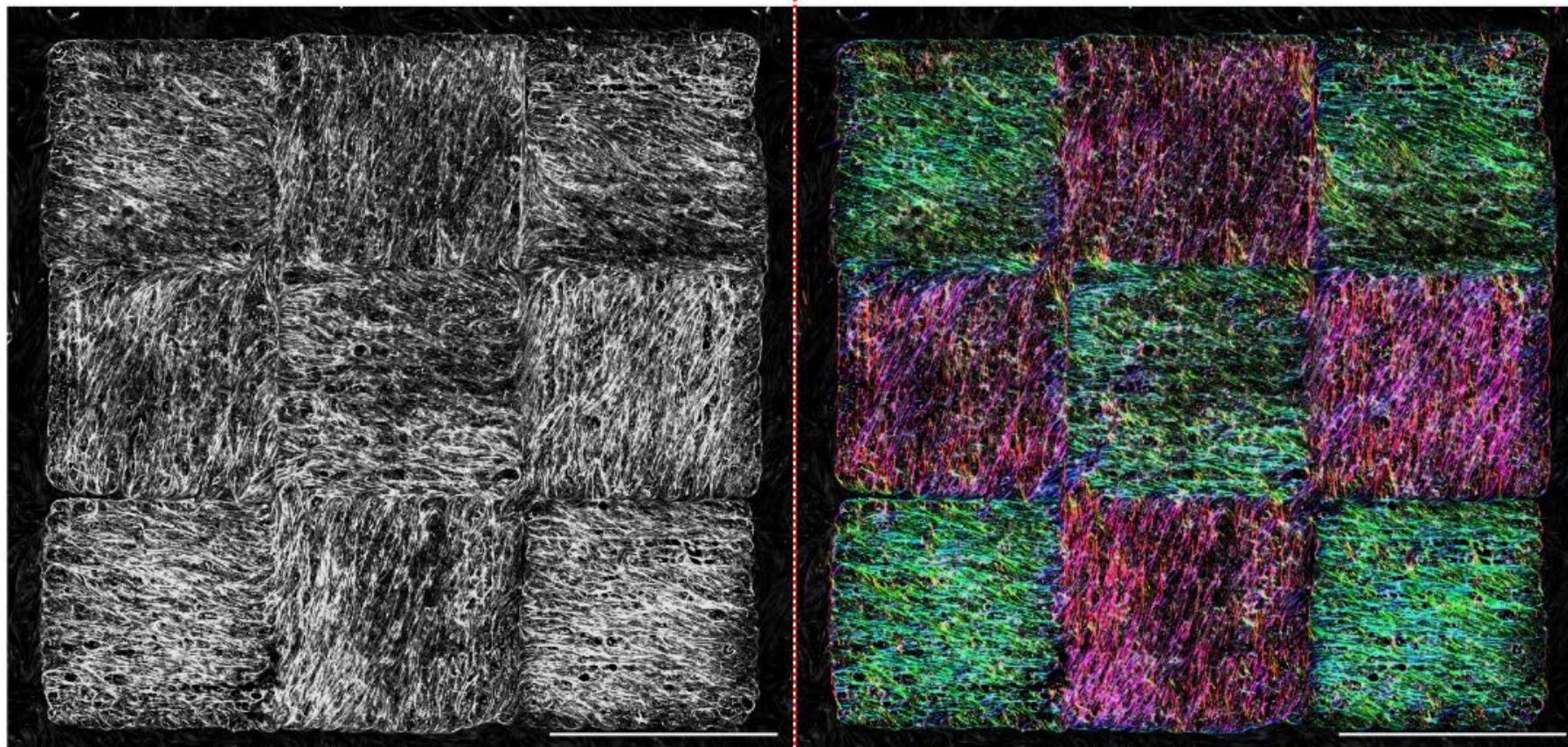
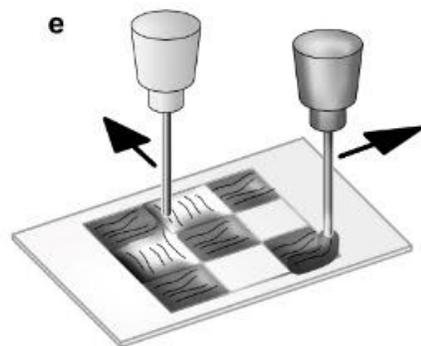
Adapted from SD Cakal et al *Biomedical materials* 2022

# Shear-induced alignment on transparent cellulose nanofiber- gelatin composites



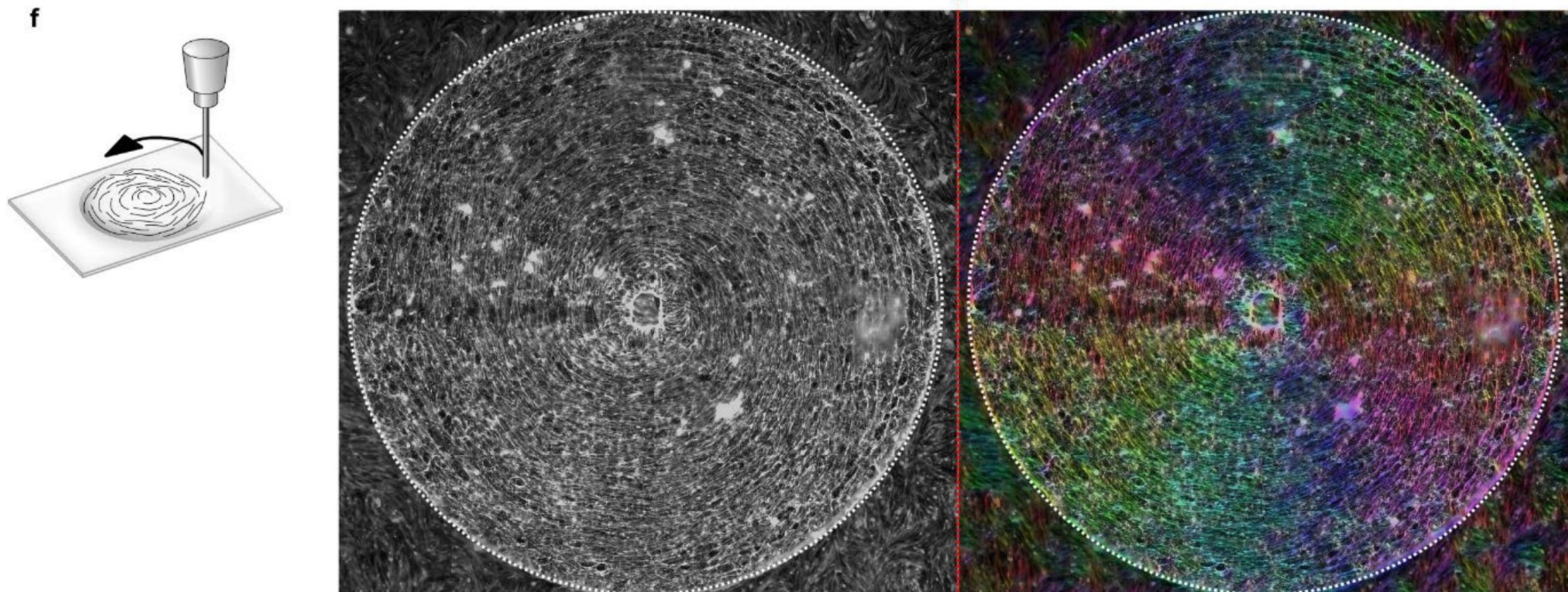
Adapted from Radeke C et al *ACS Applied Materials and Interphases* 2023

# Shear-induced alignment on transparent cellulose nanofiber- gelatin composites



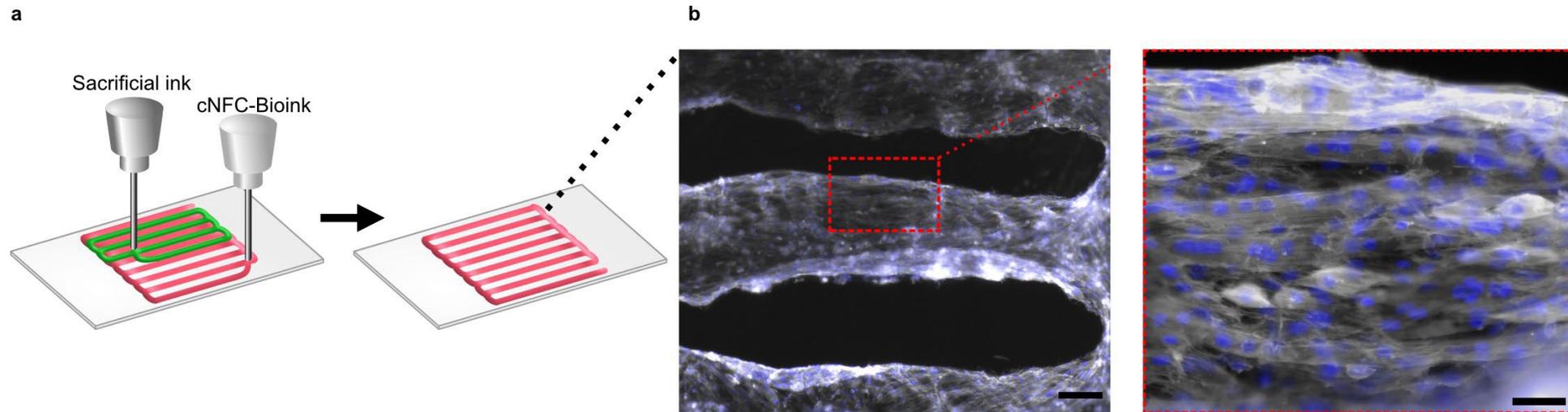
Adapted from Radeke C et al *ACS Applied Materials and Interphases* 2023

# Shear-induced alignment on transparent cellulose nanofiber- gelatin composites



Adapted from Radeke C et al *ACS Applied Materials and Interphases* 2023

# 3D bioprinted muscle fibers



# Embedded 3D bioprinting

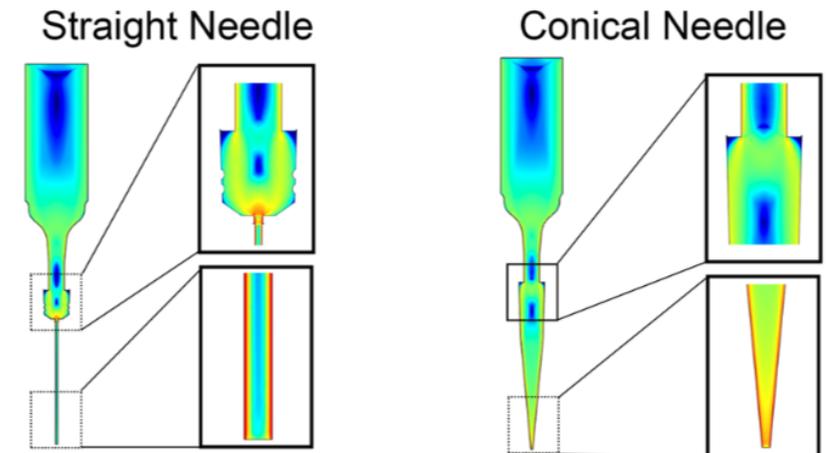
# Bioprinting: shear during print – cell viability

Shear-stress in a tube is

$$\text{Circular tube: } \tau_w = \frac{4\eta Q}{\pi R^3}$$

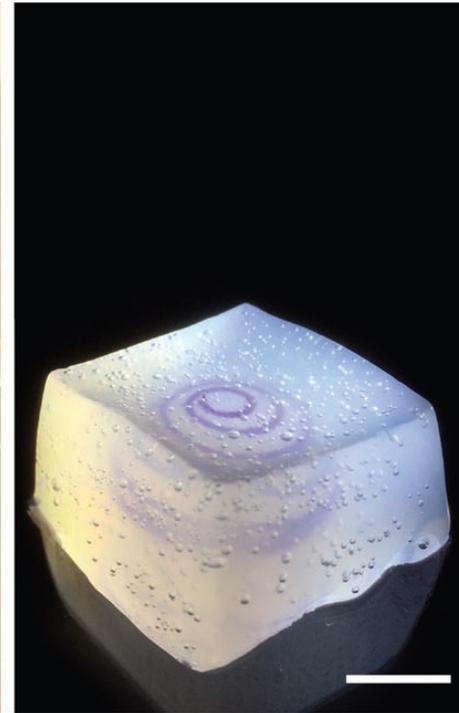
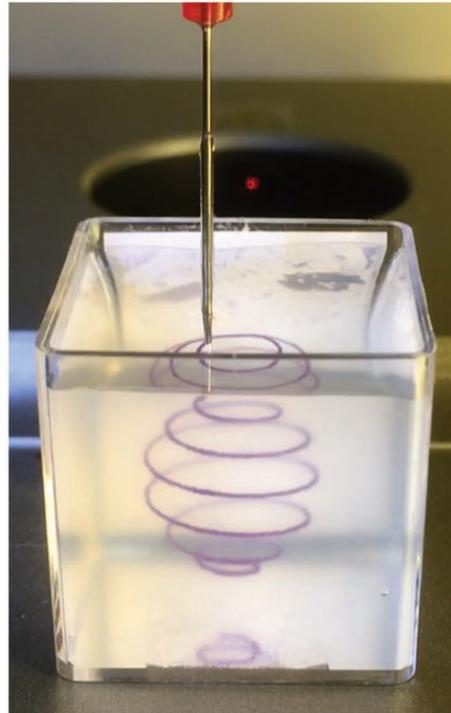
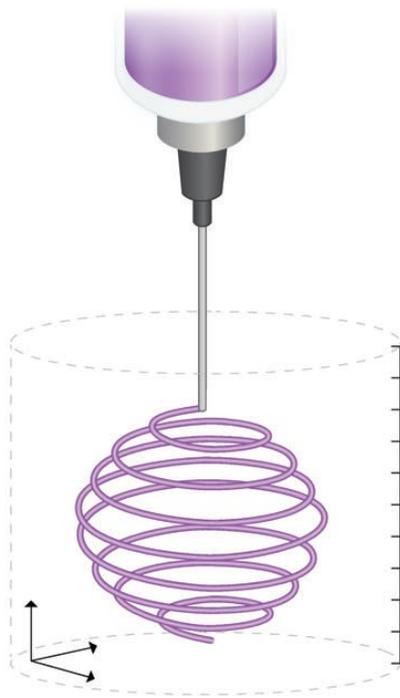
$Q$  is the volumetric flow rate,  $R$  = radius,  $\eta$  = viscosity

→ Inherent trade-off between cell viability & phenotype preservation and spatial resolution



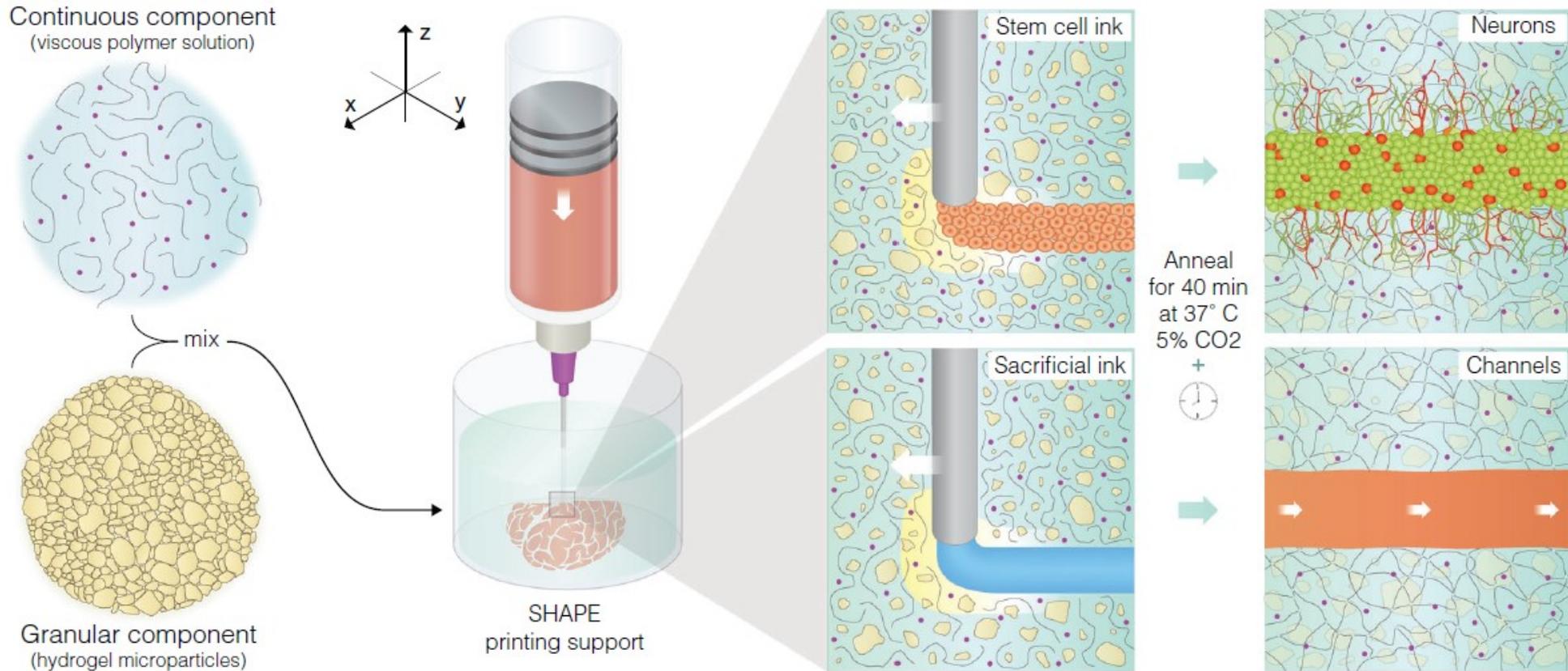
# Embedded bioprinting

- Core idea: Printing low viscosity, high-cell concentration inks into support that maintains shape



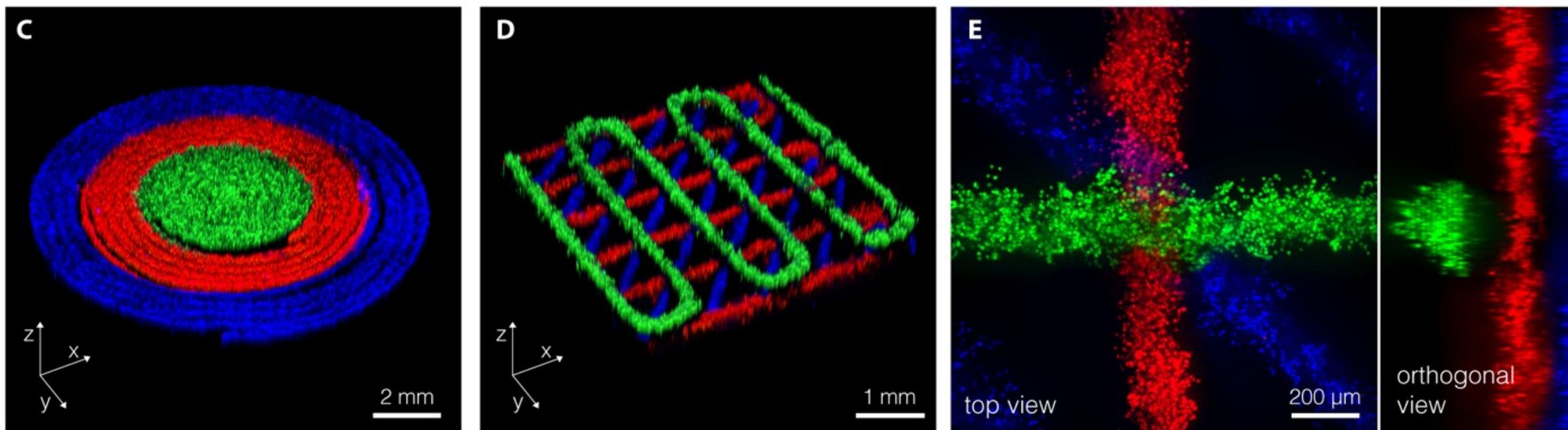
Adapted from J Kajtez et al *Advanced Science* 2022

# Embedded printing in diluted granular gels



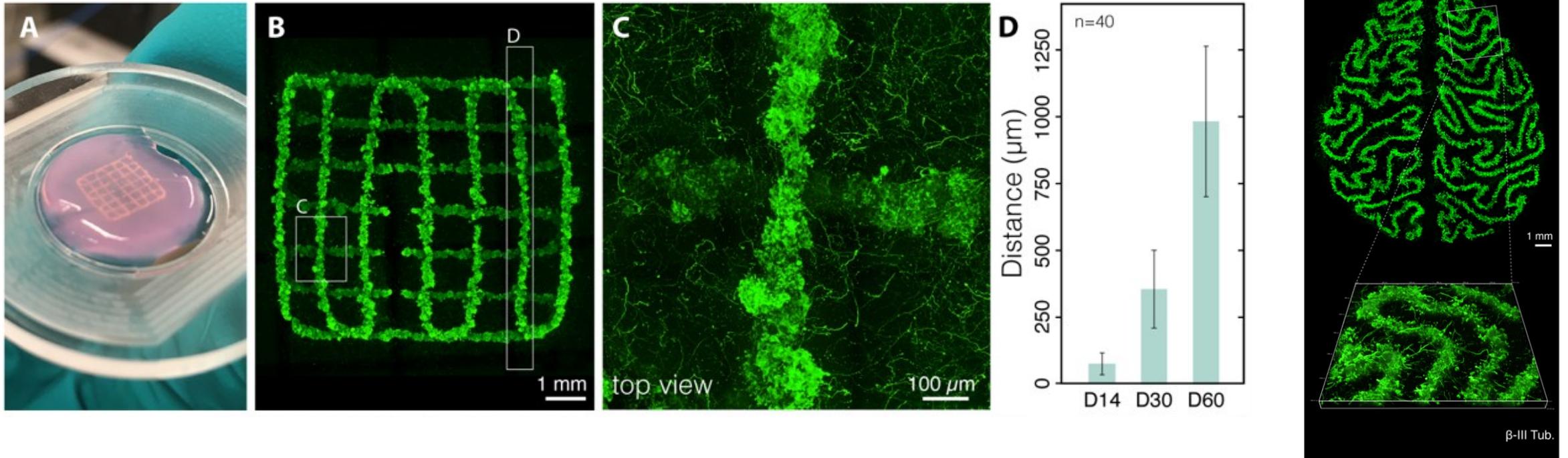
Adapted from J Kajtez et al *Advanced Science* 2022

# Embedded printing in diluted granular gels



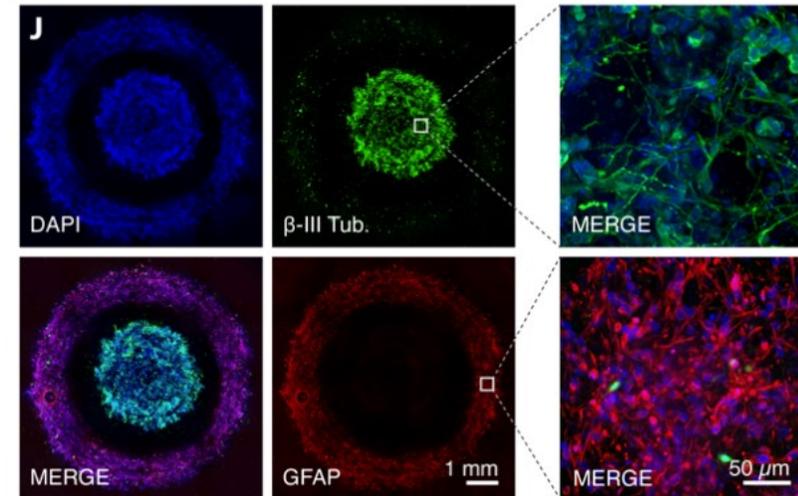
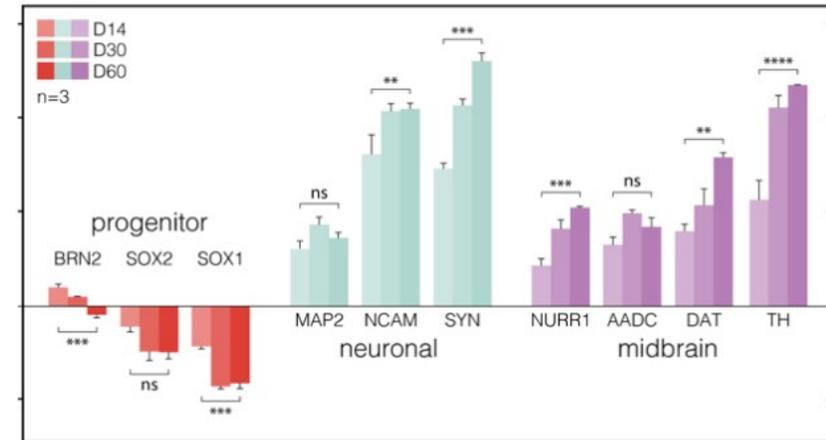
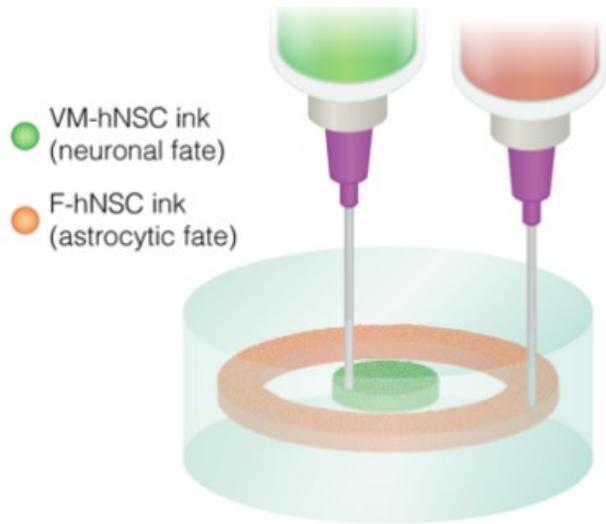
Adapted from J Kajtez et al *Advanced Science* 2022

# Embedded bioprinting of neuronal tissue models



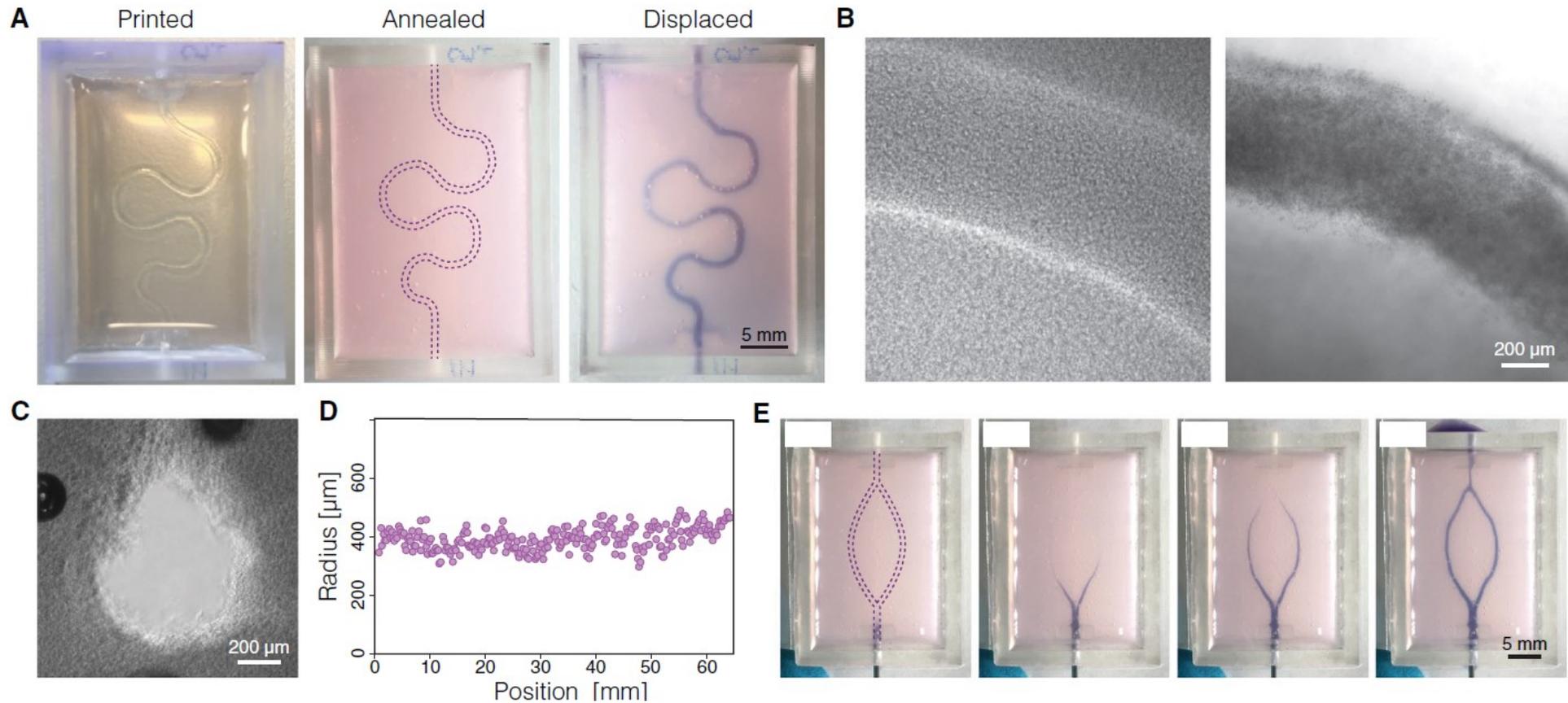
Adapted from J Kajtez et al *Advanced Science* 2022

# Embedded bioprinting of neuronal tissue models



Adapted from J Kajtez et al *Advanced Science* 2022

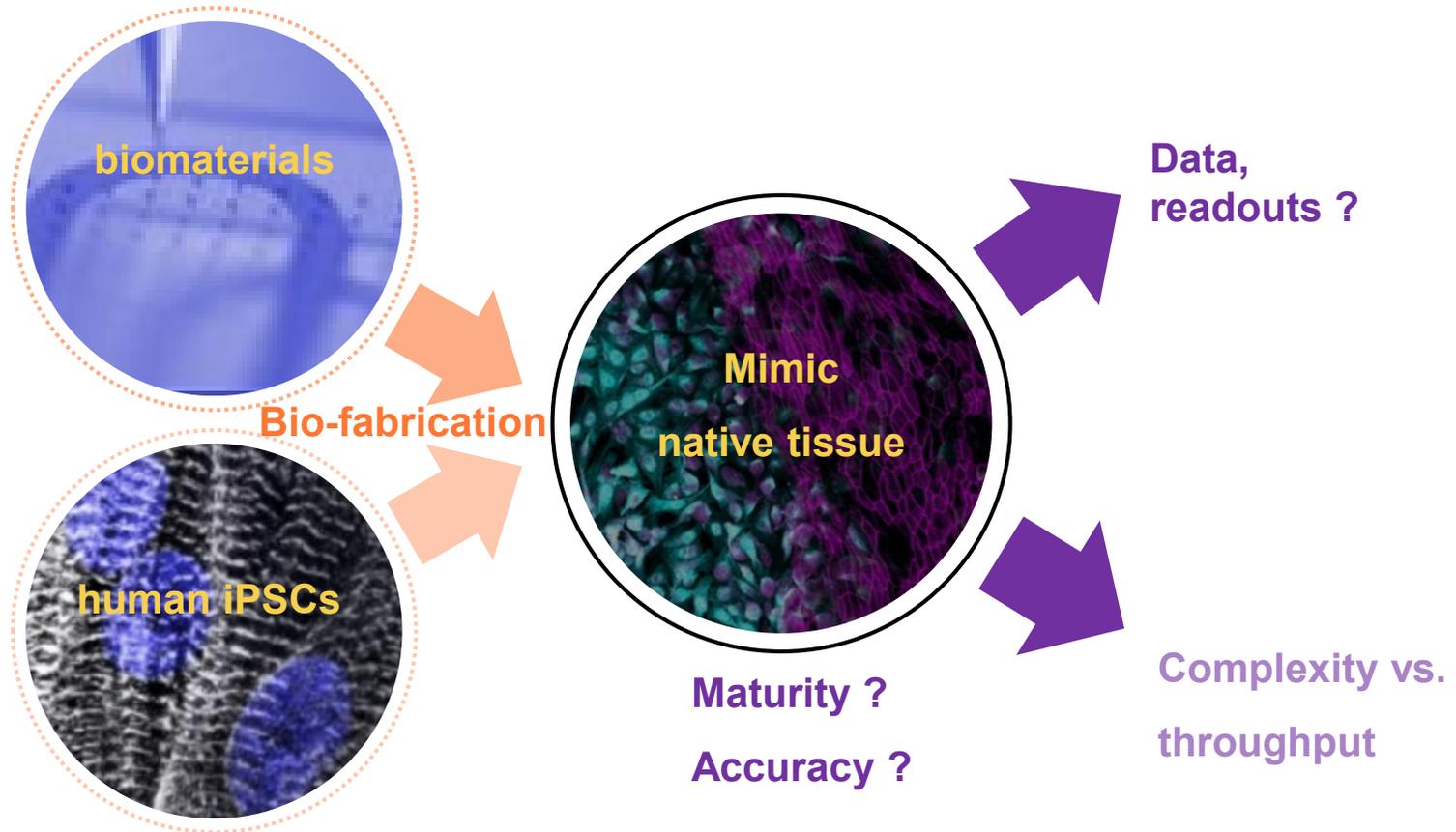
# Embedded bioprinting perfusable vasculature



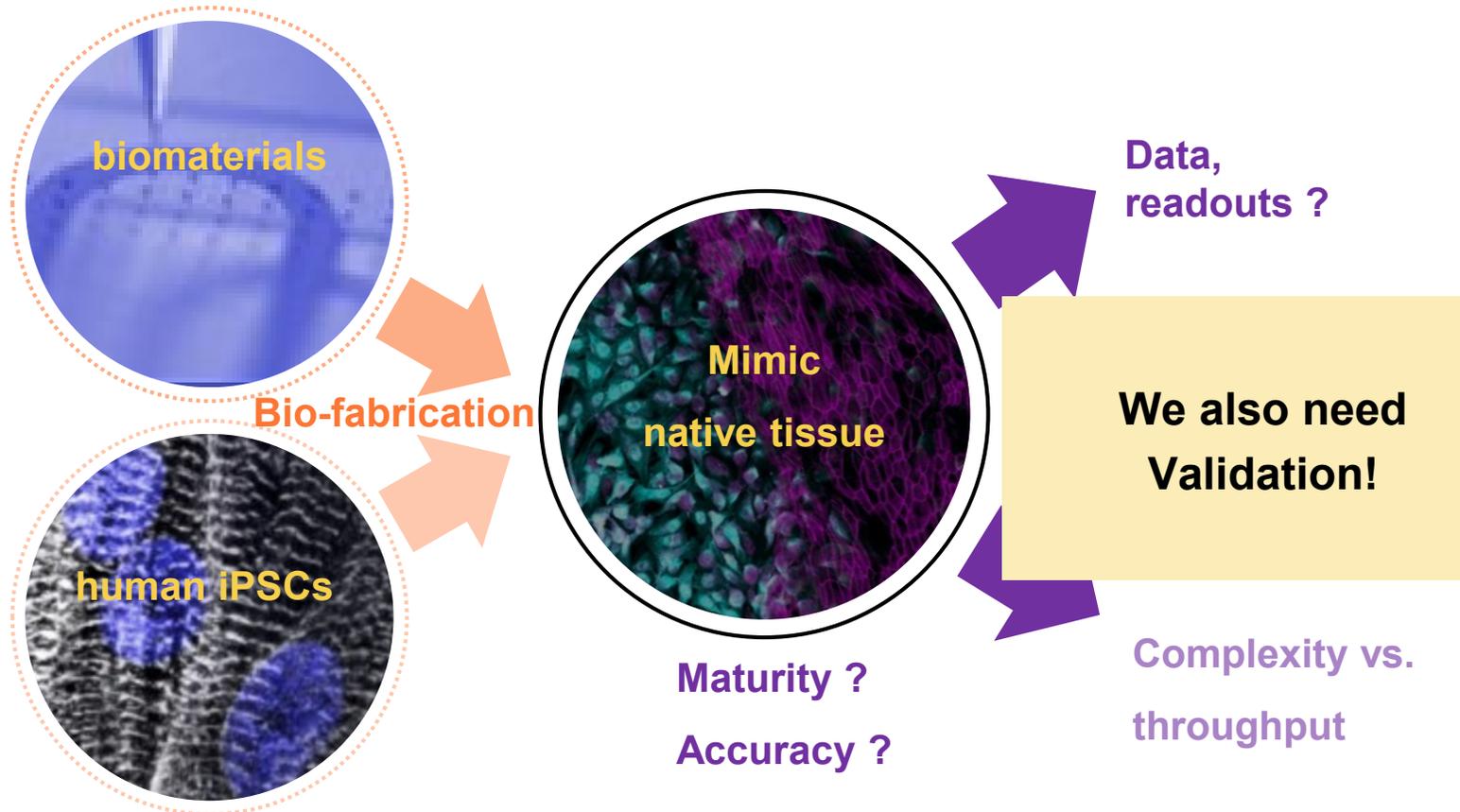
Adapted from J Kajtez et al *Advanced Science* 2022

# Summary & Outlook

# Engineering challenges

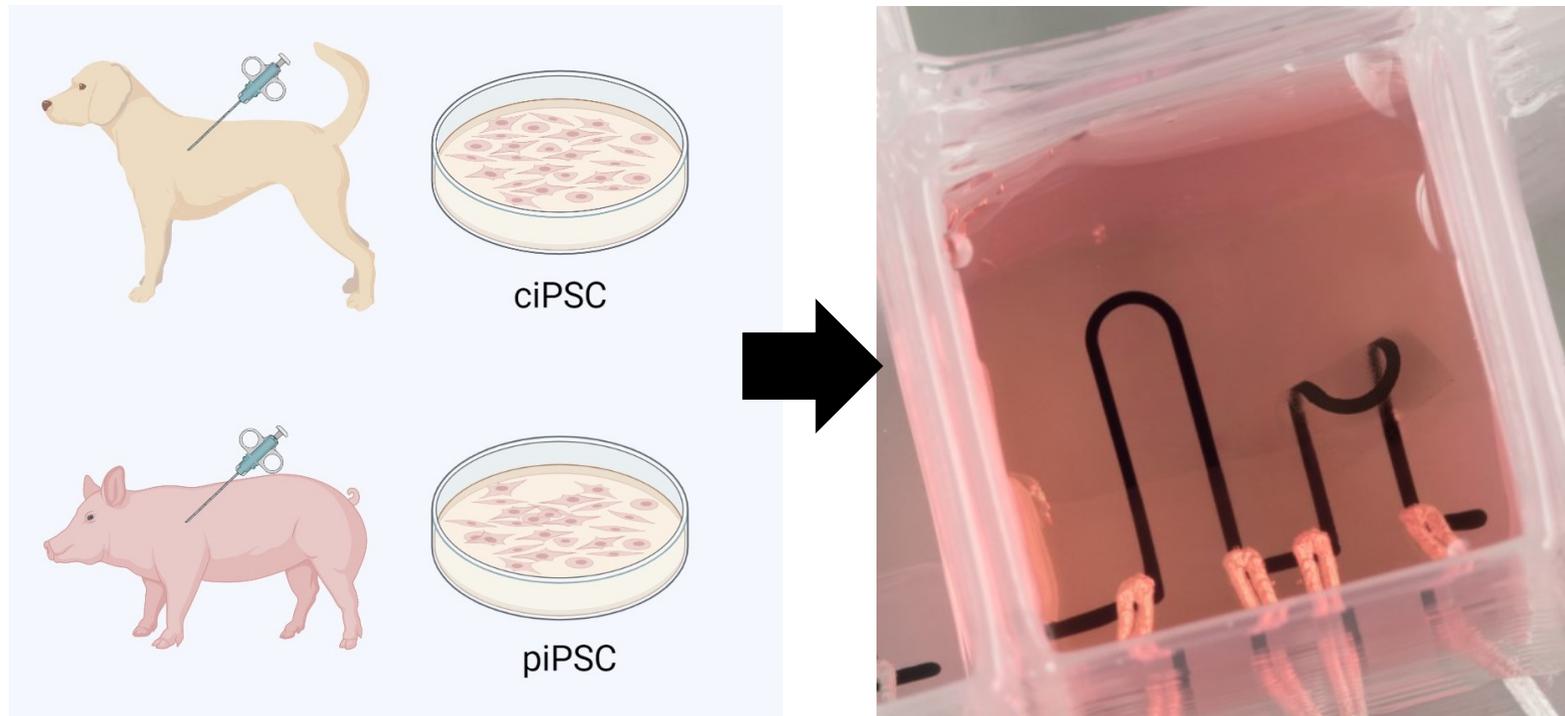


# Engineering challenges



# Validation

Historic in vivo animal data is unique opportunity for in vitro / in vivo comparisons and validations



Ongoing projects with Kirstine Callø, Kristine Freude Uni. Copenhagen to build iPSCs models from porcine and canine stem cells

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